

Follicular Unit Hair Transplantation 2010

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Summary

- Follicular unit transplantation (FUT) is a method of hair restoration surgery that relocates hair in follicular units, its naturally occurring groupings. The small size of the units allows for tremendous versatility in their placement, the creation of hair patterns that closely mimic nature and the transplantation of a large enough number of grafts so that a full restoration can often be achieved in just one or two sessions.
- Follicular unit transplantation requires large numbers of grafts, so it is important to accurately assess patients' donor reserves in the initial planning. Densitometry enables the physician to estimate the total amount of movable hair, the size of the individual follicular units, and the degree of miniaturization in both the donor and recipient areas; scalp laxity assessment is also crucial to the surgical planning.
- Great care should be taken to screen for patients who are too young or whose hair loss may be diffuse, making the donor supply inadequate for a hair transplant.
- Identifying androgenetic alopecia often proves difficult in women as other medical conditions can have a similar presentation. Options for medical treatment are more limited in women and many experience diffuse hair loss, a relative contraindication to surgery.
- Determining the right size and location of the donor strip maximizes the yield and prevents problems such as widened and/or visible scars.
- The follicular units obtained from donor strips are delicate structures. Careful dissection, gentle handling, and adequate hydration are essential to their survival. In order to dissect and place the thousands of grafts often required in FUT, the surgical team must be proficient in stereomicroscopic dissection and insertion techniques—skills that can take a year or more to develop.
- Follicular unit transplantation requires an astute aesthetic sense for optimal hairline design, correct angling and distribution of grafts and the establishment of a master plan that anticipates further hair loss and is consistent with the long-term goals of the patient.

Introduction

Within the past decade, follicular unit transplantation (FUT) has transformed hair transplantation from a cosmetically unpredictable procedure to one that can produce consistently natural results. The key to its effectiveness lies in the fact that scalp hair tends to grow in tiny bundles, called 'follicular units,' rather than individually.¹ By working with these naturally occurring units, instead of larger or smaller grafts, FUT creates as natural a look as possible while minimizing the transplant wound size and accelerating postoperative healing.

Follicular units are made up of one to four terminal hairs, one to two vellus hairs, their associated sebaceous glands, neurovascular plexus, an erector pilorum muscle, and a circumferential band of adventitial collagen, the perifolliculum.² The tendency of scalp hair to grow in this way, rather than as single hairs, can be most easily demonstrated by clipping the hair to approximately 1 mm

in length and then viewing it with a densitometer (Fig. 29.1) at $\times 30$ magnification in a 10-mm field.³ What this also reveals is that these compact units are surrounded by significant amounts of non-hair-bearing scalp (Fig. 29.2).⁴ Including this extra skin in the dissection—as do transplants with larger grafts, such as plugs and mini-grafts—requires a larger recipient wound, as well as risking visible scarring in the skin around the grafts and distortions of the growing hair.

In follicular unit transplantation these naturally occurring units are used exclusively. The FUT procedure takes advantage of the anatomic proximity of the hairs within each unit to:

- keep the recipient site wound size to a minimum
- prevent skin surface change in the recipient area
- facilitate postoperative healing
- enable grafts to be placed very close together
- permit large numbers of grafts to be safely transplanted in a single session (because of the small recipient wound size).^{1,5}

The ability to insert up to four hairs in a tiny recipient site is especially valuable cosmetically and has been instrumental in eliminating the see-through quality of micrografting. In the end, what this means is that the transplanted hair will look natural – given that the transplant surgeon makes the right aesthetic decisions about graft placement, angling, and distribution.^{6,7}

The higher precision of FUT as compared to mini-micrografting is especially obvious when methods of harvesting the grafts are compared. In FUT, once a strip of donor tissue has been removed, follicular units are isolated through careful stereomicroscopic dissection, avoiding injury to the follicles.^{8,9} The grafts in mini-micrografting, on the other hand, may or may not correspond to individual follicular units. They are cut from the donor strip according to the number of hairs they contain (optimally one to six, but often more) and/or the size of tissue required to fit into a given recipient site—a process known as ‘grafts cut to size.’ Often the dissection is performed without magnification, and it is also common for the donor tissue to be removed with a multibladed knife, rather than as a single strip, causing significant destruction of follicles. Because the grafts may be too large to fit into a slit incision, an excision (where recipient tissue is removed, such as with a small punch) is sometimes needed. In contrast, the recipient sites used for FUT always consist of small incisional slits. The advantages of FUT over mini-micrografting techniques have accounted for the shift to FUT over the past decade.

In a new procedure called follicular unit extraction (FUE),¹⁰ the need for a donor strip is eliminated entirely. In this procedure, individual follicular units are removed (extracted) directly from the donor area through a small circular incision with, or without, microdissection. Although this procedure obviates the need for a linear strip, it has limitations, particularly in the inability to efficiently utilize all the tissue in the mid-portion of the permanent zone. As of this writing, FUE is used in only a small minority of patients requiring surgical hair restoration. A description of this procedure will be included in the section on donor harvest under Techniques.

The purpose of this chapter is to describe the basic technical skills and aesthetic judgments involved in follicular unit transplantation. It should be taken as a framework for physicians interested in learning the procedure, not an exhaustive review of the various surgical techniques

used in FUT or of other hair restoration methods. The chapter will not cover mini-micrografting,¹¹ the combining of large and small grafts,¹² laser hair transplantation,^{13,14} scalp reductions,^{15,16} scalp lifts,¹⁷ or flaps.¹⁸ These procedures are infrequently used today and have been detailed in many excellent publications, as well as in two comprehensive textbooks.^{11,19}

When learning FUT, the importance of hands-on experience and beginning with small sessions until skills develop cannot be overemphasized. Having a well-trained team of assistants is also important to a successful outcome, particularly for those procedures involving a large number of grafts.

Although simple in concept, follicular unit transplantation has many nuances and complexities. Those wishing to perform FUT in their clinical practice are encouraged to join the International Society of Hair Restoration Surgery (ISHRS) and attend its annual meeting, to subscribe to *Hair Transplant Forum International* (the trade publication for hair restoration surgeons), and to follow relevant medical literature. Although it is an independent board, certification by the American Board of Hair Restoration Surgery (ABHRS) indicates a basic competency in the field. Certification requires three years of clinical experience and passing both oral and written examinations. The process of preparing for ABHRS certification is a worthwhile endeavor and recommended for those serious about surgical hair restoration.

Historical Vignette

Reports of successful hair transplants appeared as early as 1930 in the Japanese literature, beginning with Sasagawa's hair-shaft insertion procedure²⁰ and then Okuda's technique of using 2- to 4-mm punches for the treatment of various alopecias of the scalp, eyebrows, and moustache. Okuda made the important observation that using smaller punches in the recipient area improved cosmetic results.²¹

By 1943, Tamura had treated 137 cases of non-androgenetic alopecia of various etiologies using techniques very similar to modern day hair transplantation.²² For instance, he harvested donor grafts by making an elliptical incision that was sutured closed, prepared recipient sites with a thick needle, stored grafts in physiologic saline, and observed postoperative telogen effluvium (shedding). Most significantly, Tamura demonstrated that single-hair grafting resulted in growth practically indistinguishable from naturally grown hair—and much more natural looking than transplants using larger grafts. Unfortunately, it took several decades before western surgeons would apply Tamura's insights to their hair restoration procedures.

The first hair transplant in the United States was performed by Dr Norman Orentreich in 1952 with grafts measuring 6–8 mm in diameter,²³ significantly larger than those of either Tamura or Okuda. At first, incredulous editors rejected Orentreich's work, not believing that hair transplantation was even possible. In 1959, the *Annals of the New York Academy of Science* finally agreed to publish his work. The paper laid out the concept of 'donor dominance' – the idea that grafts continue to show the characteristics of the donor site after they have been transplanted to a new location. This remains the basic tenet of all hair transplantation surgery. Yet while donor dominance insured that transplanted hair could grow, it did not guarantee that the results would look natural.

Not until 40 years later would hair transplants in the United States start to produce consistently natural-looking results and promise predictable cosmetic improvements in most patients. It was a slow evolution, but the large grafts used throughout the 1960s and 1970s eventually gave way to mini-grafts in the 1980s³ and mini-micrografting in the early 1990s.²⁴ The stage was then set for follicular unit transplantation. First appearing in the medical literature in 1995¹ it quickly emerged as the standard in hair restoration—supplanting mini-micrografting in the treatment of androgenetic alopecia and rendering other well-established procedures such as scalp reductions, scalp lifts, and flaps virtually obsolete.

So swift was the ascent of FUT that the two standard textbooks on surgical hair restoration, published in 1995²⁵ and 1996,¹¹ as well as the most comprehensive text on trichology, published in 1997,²⁶ make not one mention of the terms ‘follicular unit’ or ‘follicular unit transplantation’. At the 1996 meeting of the International Society of Hair Restoration Surgeons, three 7-minute presentations on the procedure were given; but at the 2002 meeting, FUT was the subject of entire seminars and workshops and suffused every aspect of the week-long gathering.

The follicular unit was first defined by Headington in his landmark 1984 paper entitled ‘Transverse microscopic anatomy of the human scalp.’² Follicular unit transplantation had its origins in the microscopic dissection techniques of Dr. Limmer in 1988 that was described in his paper ‘Elliptical donor stereoscopically assisted micrografting as an approach to further refinement in hair transplantation’ in 1994.⁸ The term ‘follicular unit’ was introduced into the hair transplant literature by Bernstein and Rassman in 1995. The conceptual framework for FUT was mapped out by these authors in the 1995 publication ‘Follicular transplantation’¹ and in the paired articles ‘Follicular transplantation: patient evaluation and surgical planning’ and ‘The aesthetics of follicular transplantation’ in 1997.^{6,7}

The name ‘follicular unit transplantation’ was formalized by a group of hair restoration surgeons in a 1998 publication in *Dermatologic Surgery*.²⁷ In this paper, the procedure was precisely defined and included the two basic techniques, single-strip harvesting and stereomicroscopic dissection, as integral parts of the procedure. However, because follicular units can now be harvested directly from the donor area without the necessity of a strip incision (using follicular unit extraction¹⁰) the original definition has become too restrictive.

The term follicular unit transplantation should now be used to encompass all hair restoration procedures that utilize naturally occurring individual follicular units exclusively in the surgery, regardless of how these units are harvested. The caveat, of course, is that the harvesting technique must always maintain the follicular unit’s integrity.

Preoperative Preparation

Patient evaluation and surgical planning

Diagnosis and classification of androgenetic alopecia

The diagnosis of androgenetic alopecia in men is usually straightforward. It is made by observing a ‘patterned’ distribution of hair loss. It is confirmed by the presence of miniaturized hairs in the areas of thinning. The diagnosis is supported by both the inexorable progression of

the hair loss according to a recognizable pattern and a history of baldness in the family.²⁸ In women, the diagnosis is more complex since the most common presentation, a diffuse pattern, can be mimicked by a host of non-androgenetic etiologies.

Miniaturization—the progressive diminution of the hair shaft’s diameter and length in response to androgens—can be most readily observed with a densitometer,¹¹ a handheld instrument that magnifies a small area of the scalp where the hair has been clipped to about 1 mm. One type, the Rassman densitometer,³ magnifies by $\times 30$ in a visual field of 10 mm^2 , making it easy to spot miniaturization and calculate hair density (Figs 29.1–29.3). With video-densitometry (Fig 29.1b), the doctor can project the image onto a computer screen so that it is immediately visible to the patient. It can also provide a permanent digital record of the person’s density and miniaturization status.²⁹



Figure 29.1 (A) Hair densitometer. (B) Video-densitometer.



Figure 29.2 Natural hair groupings as seen through a densitometer (magnification $\times 30$).

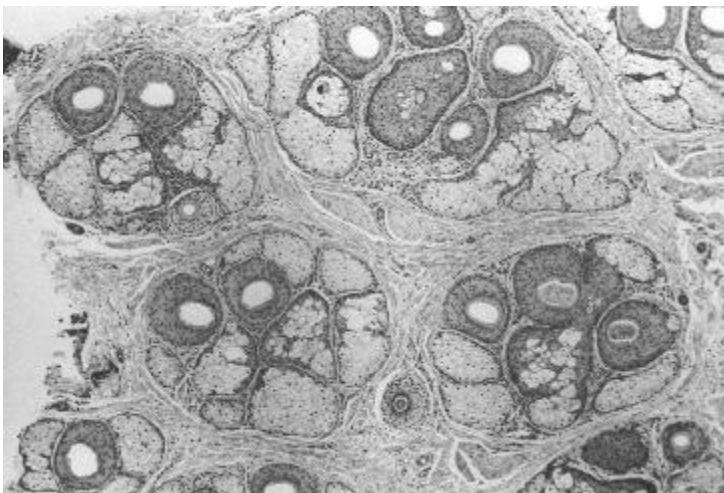


Figure 29.3 Histological view of follicular units corresponding to the natural hair groups observed on the scalp surface. Transverse section taken at the level of sebaceous glands (hematoxylin and eosin; original magnification $\times 40$).^{2,5}

When describing hair loss, particularly in the early phases, it is preferable to think in terms of changes in volume rather than density. Density is simply the number of hairs per unit area, whereas volume reflects both hair shaft diameter as well as the absolute numbers of hairs.³⁰ A new device, called the Trichometer, takes both of these factors into account by measuring the cross sectional area of a demarcated bundle of hair.³¹ The earliest sign of hair loss, that of miniaturization, results in volume changes caused by the individual hair decreasing in size without a decrease in their number. Only in more advanced balding will the actual number of hairs start to decrease.¹ This is why devices that can assess hair shaft diameter are so useful for identifying early hair loss.

The Norwood classification of male pattern hair loss, published in 1975, remains the most widely used.³² It defines two major patterns and several less common types (Fig. 29.4). In the regular Norwood pattern, two areas of hair loss—a bitemporal recession and thinning crown—gradually enlarge and coalesce until the entire front, top, and crown (vertex) of the scalp are bald. A less common Norwood variety, Norwood class A, is characterized by a distinctly anterior-to-

posterior progression, usually resulting in baldness on the front and top of the scalp, but more limited loss in the crown. In both patterns, the sides and back tend to resist androgenetic changes, though the sides may exhibit significant thinning in senile alopecia.



Figure 29.4 Norwood classification.³²

Two other types of genetic hair loss not emphasized in the literature, diffuse patterned and diffuse unpatterned alopecia, pose the greatest challenge both for diagnosis and patient management. A thorough familiarity with all of these patterns is essential for diagnosing androgenetic alopecia and planning FUT (Table 29.1).

Table 29.1 Classification of androgenetic alopecia	
Norwood classes	Regular classes I to VII
	Type A variant (IIa to Va)
	Variations: persistent frontal forelock ⁹⁷ ; persistent frontal hairline
Diffuse androgenic alopecias	Diffuse patterned alopecia (DPA)
	Diffuse unpatterned alopecia (DUPA)
Senile alopecia	
Female pattern alopecia	Ludwig classes I – III

Diffuse patterned alopecia (DPA) is an androgenetic alopecia manifested as diffuse thinning in the front, top, and vertex, with a stable permanent zone. In DPA, the entire top of the scalp gradually miniaturizes without passing through the typical Norwood stages. Diffuse unpatterned alopecia (DUPA) is also androgenetic but lacks a stable permanent zone and affects men much less often than DPA. DUPA tends to advance faster than DPA and eventuates in a horseshoe pattern resembling the Norwood class VII. However, unlike the Norwood class VII, the DUPA horseshoe can look almost transparent due to the low density of the back and sides. Differentiating between DPA and DUPA is very important because DPA patients often make good transplant candidates, whereas DUPA patients almost never do, as they inevitably suffer extensive hair loss without a stable zone for harvesting.⁶

The early warning signs of DUPA include:

- A rapid decrease in hair volume and a change in hair texture at an early age, often in the teens.
- The maintenance of an adolescent hair pattern and persistent frontal hairline in spite of dramatic volume change.
- A see-through donor area (most easily visualized when the hair is wet or lifted up)
- Abnormal Densitometry
 - Significant miniaturization of the donor area (greater than 35%)
 - A donor density of less than 1.5 hairs per mm²

The sensitivity of Densitometry in detecting early DUPA justifies its routine use in the evaluation of hair loss. If a diagnosis of DUPA is even remotely suspected, any decisions regarding surgical hair restoration should be postponed. The possibility of missing a DUPA diagnosis is one of the most powerful arguments against performing hair transplantation at an early age.

Both DPA and DUPA occur in women, but—in contrast to men—DUPA is far more common. As with men, women with DUPA do not make good transplant candidates (except possibly when donor hair is used solely to soften the frontal edge of a wig). Indeed, the higher DUPA incidence in women explains why a smaller proportion of women than men qualify as transplant candidates. Interestingly, the most common classification used for women, the Ludwig Classification, does not differentiate between DPA and DUPA.³³

Recently, a home test has been developed that screens for variations in the androgen receptor gene on the X chromosome, a gene associated with male pattern hair loss. The purpose is to identify persons at increased risk of developing hair loss before it is clinically apparent so that medical intervention can be started early, when it is most effective. The presence of one of the variants indicates a moderately increased risk for the development of androgenetic alopecia and the other indicates a substantially decreased risk of becoming bald. A similar screening test is also available for the detection of hair loss in women.³⁴

It is important to realize that, at this point, these markers just reveal an association with hair loss and that a cause and effect relationship has not been proven. A danger is that patients may overreact to this relatively incomplete information. It is best to have the tests performed under a doctor's supervision so that the results can be put in the context of other information that the

physician gleans through a careful history and physical and so that the patient may be properly counseled.

It is important to emphasize that because a wide variety of medical conditions can produce diffuse hair loss, a non-androgenetic differential must be considered in all unpatterned alopecias. This is particularly relevant in evaluating women in whom unpatterned hair loss is the rule rather than the exception (Table 29.2). The following laboratory tests are often useful when a non-androgenetic cause for diffuse hair loss is suspected: chemistry screen, complete blood count, serum iron, total iron-binding capacity, triiodothyronine, thyroxine, thyroid-stimulating hormone, antinuclear antibody, and serologic test for syphilis.

Table 29.2 Non-androgenetic causes of diffuse hair loss in women
Anemia
Endocrine diseases (especially thyroid)
Connective tissue disease
Obstetric/gynecologic conditions (e.g. postpartum, polycystic ovarian disease)
Weight loss (esp. crash diets)
Emotional and physical stress (e.g. surgery, anesthesia)
Medications: oral contraceptives, thyroid drugs, anti-hypertensives (especially β -blockers), psychotropics, anticoagulants, anti-lipemics, gout therapy, prednisone, excessive vitamin A or tryptophan use

In women, diffuse or patterned hair loss may be a sign of excessive androgen production. A further medical evaluation is indicated when the hair loss is associated with any of the following: cystic acne, irregular menses, hirsutism, virilization, infertility, or galactorrhea. Serum levels of DHEA-sulfate, androstenedione, total and free testosterone, and prolactin can serve as a useful screen.³⁵

When the diagnosis of androgenetic alopecia is uncertain, further diagnostic information can be gleaned from a hair-pull test for telogen effluvium, a potassium hydroxide mount and culture for fungus, a microscopic examination of the hair bulb and shaft, and a scalp biopsy (sectioned horizontally).³⁶ A dermatologic consultation is warranted whenever the diagnosis of hair loss is unclear.

Who is a candidate for hair transplantation?

In all cosmetic procedures, a successful outcome depends on proper patient selection (Table 29.3). In surgical hair restoration, proper timing is also crucial. There are no medical or surgical advantages to transplanting at an early age. The popular rationale that transplants should be performed at a young age so that ‘no one will notice’ does not make sense: if there is nothing to notice after the transplant matures, why perform it in the first place? FUTs heal so quickly and the hair growth is so gradual that, once the short postoperative period ends, the process is almost always undetectable. When hair loss is early, medical therapy is the treatment of choice.

Table 29.3 Surgical patient selection for androgenetic alopecia
Age: 25 years or older
Inadequate response to medication after 1 year
Significant hair loss: Norwood class III or greater
Diffuse patterned alopecia (DUPA) has been ruled out
Non-androgenetic causes of hair loss have been ruled out*
No medical contraindications to surgery (drug sensitivities, keloids, connective tissue disease) [†]
Reasonable patient expectations
* <i>Many non-androgenetic causes of hair loss can be treated with hair transplantation.</i>
[†] <i>Some are relative contraindications.</i>

Patients seeking hair transplantation while still in their early 20s invariably wish to have their adolescent hairline and original density restored. Since neither of these goals can be achieved surgically without compromising the patient’s future appearance, the procedure should not be performed at this age. It should be explained that hair transplantation is a procedure that moves, rather than creates, hair. It does not increase overall hair volume. In fact, the challenge is to have the pattern of the transplanted hair take into account the person’s ever decreasing total hair mass. In other words, patients who will have significant balding need to have built-in temporal recession, and possibly a crown that is significantly thinner than the surrounding hair, for the transplant to look natural over time. Unfortunately, neither of these is generally acceptable to a young man in his early 20s.

Another reason to postpone transplants for those under 25 years, or with only limited hair loss, is to give medications—particularly finasteride—adequate time to work. As patients may continue to show regrowth for 1 to 2 years after treatment is initiated, they should in general take finasteride at least this long before surgery is considered. It is inappropriate, therefore, to start younger patients on finasteride and schedule surgery at the same time. Fortunately, finasteride works best in younger patients, and especially those with large areas of miniaturized scalp, rather than areas that are totally bald. On the other hand, for older patients who would be taking finasteride to maintain rather than regrow hair, and for those whose advanced state of hair loss makes results from finasteride unlikely, the medication may be prescribed concomitantly with surgery (see Preoperative Preparation).

With the availability of finasteride, medication plays an increasingly important role in the management of androgenetic hair loss. Although the incidence is small (2-4%), the stigma of possible sexual side effects has turned many against this drug. Some also consider surgery a more conservative form of treatment than the potential lifetime use of a drug. Because of these concerns and the pressure from panicked patients for surgery’s apparent “quick fix,” the physician should take time to thoroughly discuss the pros and cons of medical therapy.

In the Norwood Classification, class I represents a normal adolescent pattern and class II a normal non-balding adult. Therefore, as a minimum, patients should qualify as class III before

transplants are contemplated. Early class III patients will often benefit from medication alone, so this should be considered first.

Although the opportunities for a complete restoration in a person who is extensively bald (ie., a Norwood class 7) is limited, extensive balding is not necessarily a contra-indication to surgery. If the person's donor zone is stable (miniaturization of less than 20%) and the patient's expectations are commensurate with his or her limited available donor supply, a hair transplant may be considered. As long as the patient is in good health, there is no upper age limit for surgery. In fact, older patients tend to have the most reasonable expectations and are often the most satisfied. Even a small amount of hair can make a dramatic improvement in the appearance of a person who has been bald for many years. .

There are few absolute medical contraindications to surgical hair restoration, particularly because it is an outpatient procedure not requiring general anesthesia. Relative contraindications include: bleeding disorders, drugs that increase bleeding such as clopidogril (Plavix), immunodeficiency, unstable arrhythmias, chronic obstructive pulmonary disease, sensitivity to local anesthetics or epinephrine (Adrenaline), a history of keloid formation or connective tissue disease, and major psychiatric disorders. When in doubt about the patient's medical condition it is always best to get medical clearance from their primary physician.

Patients should be able to tolerate being in the surgical chair for a good part of the day; consequently, back and neck problems as well as claustrophobia can sometimes make the procedure problematic. Special care should be taken when evaluating patients with significant psychiatric problems, particularly depression, trichotillomania, body dysmorphic disorder, or severe OCD. In such cases, a psychiatrist or psychologist should participate in the decision-making process.

Possibly the most difficult aspect of the consultation is managing patients' expectations. It is almost a cliché to say that they must have 'reasonable expectations', but ensuring this is the essence of the evaluation. The surgeon cannot stress strongly enough that, because hair restoration procedures move rather than create hair, the resulting transplanted density will be significantly less than the person's non-balding density. It is also important to emphasize that it is the patient's own hair from the sides and back of the head that will be transplanted. Therefore, the final result can be best described with reference to the texture, color, curl, and other qualities of the person's existing hair. In addition to a thorough discussion, it is also helpful to provide prospective patients with printed material and photos, and to make past patients available for them to meet.

Assessing the patient's donor supply

The main factors in determining total donor reserves are donor density, scalp laxity, and the physical size of the donor area.

Donor density

The number of follicular units in the mid-portion of the donor area of a normal Caucasian man who has not had surgery is approximately one unit per mm^2 .⁷ In the first procedure, therefore, the donor area should yield approximately one follicular graft per mm^2 of scalp, provided there is no significant wastage during surgery. The average yield declines when longer strips are taken because density decreases towards the sides. In some people, density varies dramatically throughout the donor area, but this is the exception rather than the rule.

Since the number of follicular units per unit area stays relatively constant, donor density is an indication of the average number of hairs per graft. For example, if a patient has a density of two hairs per mm^2 , the donor area will generally contain one follicular unit per mm^2 and the grafts will consist of a combination of one-, two- and three-hair units, with the average being two hairs per graft. With a density of 2.3 hairs per mm^2 , there will still be one follicular unit per mm^2 , so the same number of grafts will be harvested per unit donor area, but the grafts will have an average of 2.3 hairs per mm^2 and consist of a mix of fewer one-hair units and a larger percentage of three- and four-hair units.⁷

Follicular unit density varies according to race, therefore the number of grafts obtained per unit area of donor tissue will exhibit racial variability and must be accounted for in the surgical planning.⁶

If the scalp has been stretched from previous transplants, scalp reductions, or scalp lifts, the follicular units will be spaced further apart, making it necessary to actually measure the density of follicular units to accurately estimate the number of grafts obtainable from the strip (as the density of follicular units will now be less than one per mm^2).

Donor scarring from previous surgeries will also significantly diminish the donor yield. All donor harvests, no matter how perfectly executed, entail some loss of hair. In addition, the angle of the hair surrounding the scar will be altered slightly, creating more transection in any subsequent harvest.

A person can lose a substantial amount of hair volume—either due to actual loss of hair or to miniaturization—before it becomes noticeable. When the hair is blonde or white it takes longer for thinning to show, while it is evident sooner for those with black hair and white skin. For those with average density and average hair attributes, approximately half of the hair in the donor area may be moved without a significant change in appearance.

Scalp laxity

Scalp laxity (looseness) is an important factor in determining a person's total available donor hair. There appears to be two main factors that contribute to laxity; redundancy and distensibility. Redundancy is simply extra scalp skin. Distensibility is the ability of the scalp to stretch. In those with lax scalps due to redundancy, harvesting the donor strip merely removes some of the extra scalp tissue while only slightly affecting density (hairs per mm^2); therefore, the yield can be high over several procedures. With little redundancy, however, each procedure

stretches the skin, measurably decreasing density. The limitations of a tight scalp are usually not apparent in the initial surgery, but in subsequent procedures it can become much harder to perform a non-tension closure or, in the face of decreased density, to harvest a significant amount of hair. Therefore, the long-term goals of those with tight scalps must reflect a more limited donor supply.

Patients with very loose scalps, due to increased distensibility, tend to heal with widened scars and so often make poor candidates for procedures using a linear incision. Some, in fact, may have undiagnosed connective tissue disease.³⁷ Follicular unit extraction should be considered in cases of both very loose and very tight scalps.¹⁰

Donor dimensions

The mid-portion of the harvestable donor zone generally lies over the occipital protuberance and extends laterally to within 3 cm of the temple hairline on either side. This distance is approximately 32–34 cm. Recession at the temples can signal extensive balding and portends a limited donor supply. The height of the permanent zone can vary markedly from person to person and should be carefully measured. Hair should be harvested only where it is stable, that is, where it lacks significant miniaturization. An often overlooked sign is the ‘ascending’ hairline, characterized by miniaturization in the lower margin of the permanent zone. Both receding temples and an ascending posterior hairline indicate a shrinking donor zone and mandate conservative surgical planning.

Planning the first hair transplant session

Ideally, the main goals of the first transplant session should be:

- to establish the frontal hairline and frame the face
- to provide coverage to all bald areas of the scalp, from the frontal hairline to the vertex transition point³⁸
- to create sufficient density so that the results will look natural after one session, though additional sessions may be desired.⁶

These goals should be the ultimate aim of those performing hair restoration surgery, but may not be achievable with a small or inexperienced surgical team. In this author’s opinion, there is little medical or aesthetic justification for performing the surgery in arbitrarily small sessions. It is preferable that each procedure cover the entire area of hair loss intended to be treated and be designed to ‘stand alone.’^{6,7}

In an alternate approach to surgical planning, the objective is to achieve final density in a one-pass procedure by creating high density in a more limited area and then transplanting another area in a subsequent session. This is accomplished in part by using very small recipient sites, limiting graft size to three hairs and using the stick-and-place method, in which grafts are inserted as soon as the recipient sites are made.³⁹

As the number of grafts placed per unit area (density) rises, so too does the risk of vascular compromise that may result in suboptimal graft growth. Technical problems of popping that may cause the grafts to become desiccated or that expose the grafts to mechanical injury on reinsertion, also become more likely. Proper patient selection and technical expertise help avoid such problems, but the risks are increased nevertheless. It certainly behoves those with more limited experience to be conservative when determining the number of grafts to be transplanted.

Because the blood supply to the scalp is extensively collateralized,⁴⁰ the risk of vascular compromise is related more to the density of grafts in a specific area and the size of the recipient wounds, than the absolute numbers of grafts placed. For this reason, the transplantation of a large number of grafts over a broad area does not seem to pose the same problems as producing very high densities. In addition, popping becomes less of a technical issue when the same number of grafts are placed over a larger area. However, transplanting large number of grafts (in ‘mega-sessions’) poses its own challenges, such as increasing the time the grafts remain outside the body, requiring more staff, contributing to patient and staff fatigue, and creating organizational issues. As with dense packing, the use of very large sessions should be reserved for only the most experienced surgical teams.

Though the amount of hair needed to cover the front and top of the patient’s scalp will vary, an attempt should be made to achieve this coverage, if only lightly, in the first session. Crown coverage should not be a goal of the first session unless the patient has an above-average donor supply; if it is attempted in the first session the patient’s options will be more limited and the chances for an aesthetically balanced transplant may be permanently compromised.

Table 29.4 offers general guidelines as to the number of follicular units recommended for the first hair transplant procedure.

Table 29.4 Number of grafts in first follicular unit transplant session		
<u>Norwood Class</u>	<u>Follicular Units</u>	<u>Total Units with Crown</u>
III	800 – 1200+	—
III Vertex	800 – 1200+	1200 – 1800+
IIIa	1300 – 1700+	—
IV	1500 – 1800+	1800 – 2200+
IVa	1700 – 2100+	—
V	1500 – 1900+	1900 – 2400+
Va	2000 – 2400+	—
VI	2000 – 2400+	2400+
VII	2400 – 2800+	2800+

Planning a second session

Timing of the second session

Transplanted hair sheds around 2–6 weeks after the procedure and the first signs of new growth occur on average in approximately 10 weeks, though the onset can vary considerably, taking as long as 4–6 months or more. Hair gradually increases in both thickness and in length, so that the initial growth is often not indicative of the final result. In less than 5% of patients, hair grows unabated after surgery, without shedding.

One should generally wait 10–12 months to best appreciate the cosmetic impact of the procedure. During this time, the continued increase in the hair's diameter and length strikingly alters the appearance of the restoration. Once the hair reaches styling length, both the patient and physician can make aesthetic judgments about the desirability of an additional treatment session.

It will also be advantageous to delay the second procedure to maximize the donor harvest. Although each procedure results in a potentially tighter scalp, some of the preoperative donor laxity returns in the months following the surgery. Although, the major change will occur during the first month as the edema and inflammation subside, further loosening occurs as the scalp stretches over the next 12 months.

In the uncommon event of telogen effluvium occurring in the donor area, the telogen follicles may not be easily identifiable in the dissection. Since recovery in the donor area may take up to a year, it is essential that patients wait until complete re-growth has taken place before a second session is performed. In rare cases, if the closure is too tight, the effluvium may eventuate in permanent hair loss.

Goals for the second session

The main goals of the second session are:

- to add density to areas transplanted in the first session
- to refine the hairline
- to follow the progression of the hair loss (if necessary)
- to extend the transplant into the crown (when appropriate)
- to complement a specific grooming pattern

In general, most patients require two sessions to achieve adequate density. When seen at one year follow-up, approximately 50% of patients are satisfied with the density achieved in the first session. However, if the first session is planned as a stand-alone procedure, the patient that does want more density does not need to have this second session in order to look natural. Of the 50% that do want further density, many postpone the next procedure due to either financial or time constraints and have it performed at a time that is convenient for them.

The hairline often can benefit from a little “tweaking” to make the leading edge softer and more irregular. Placing the hairline too low is a common mistake, but so is placing it too high, something often done with the intention of bringing it down on the second session, once the

patient can see how it looks. It is better to try to get the hairline position right in the first session; lowering it later often leaves the leading edge too thin.

The decision regarding crown coverage is important because it is the least visible of the balding regions, but can potentially occupy a very large surface area, producing an almost inexhaustible demand upon the donor supply.

If extensive balding appears likely and the patient has a modest donor supply, the crown should be treated as an extension of the top, rather than an isolated region. This will help to ensure that the patient will not be short of hair if the intervening bridge between the front and crown were to require additional grafts. Unless the patient's history, age, and physical examination indicate limited balding, it is particularly important to avoid creating high density in the crown, as subsequent balding could leave an isolated island of hair.

A useful alternative to covering the crown with transplanted hair is to end the transplant at the vertex transition point.^{38,41} The patient can then groom the hair back to conceal the non-transplanted area. This is recommended when direct crown coverage is not realistic or it is too early in the balding process to determine whether significant crown coverage will be possible in the future. Another advantage of sparing, or lightly covering, the crown is that donor reserves may be saved to address further diminution of the lateral donor fringe.

Particularly when a person has been bald for many years, it is not known how the person will want to groom his/her hair after the restoration. Once the first procedure has grown in, however, many people will feel comfortable committing to a specific grooming pattern. If this is the case, then the second procedure may be used to complement a specific way of styling the hair. For example, if a man is relatively certain that he will continue to comb his hair from left to right, then the transplant can be left-weighted to enhance the appearance of density when the hair is groomed in this manner. If, however, the person prefers to comb his hair straight back, or wants to vary his look, then the second transplant should be symmetrical like the distribution of the first.

Subsequent transplant sessions

The surgeon should make every attempt to accomplish the restoration in as few sessions as possible, rather than putting patients through an unnecessarily protracted course of multiple surgeries that can adversely affect their donor area and lifestyle. The number of grafts required to achieve patient satisfaction varies widely due to the great variability in hair characteristics. Moreover, because hair loss patterns form a continuum, while the Norwood classes are discrete, even the transplant size necessary for each class can vary significantly. Table 29.5 shows the approximate number of follicular unit grafts necessary for a complete restoration, without and with crown coverage.

<u>Norwood Class</u>	<u>Follicular Units</u>	<u>Total Units with Crown</u>
III	1000 – 1600	—
III Vertex	1000 – 1600	1600 – 2200
IIIa	1400 – 2200	—
IV	1400 – 2200	2200 – 3000
IVa	1800 – 3600	—
V	2000 – 3000	3000 – 4200
Va	2400 – 4400	—
VI	2200 – 4600	4200 – 5600
VII	2200 – 4800	4400 – 6600

The patient

Doctors commonly place patients on finasteride before surgery to minimize the chance of postsurgical effluvium (shedding). Although there are no scientific studies confirming that it is effective for this purpose, it would seem that if the goal is to prevent postsurgical shedding, the medication should be prescribed at least a month before the procedure. If, however, the intent is to postpone or obviate the need for surgery, then it should be taken for a minimum of one year to allow sufficient time for the patient’s hair to regrow.

Patients using topical minoxidil are advised to discontinue its use several days before surgery (because of its vasodilator properties) and wait until a week after the procedure before resuming (to avoid the irritating effects of the alcohol in the 2% solution or the propylene glycol in the 5% solution). Although we suggest minoxidil in combination with finasteride for patients with early hair loss, we generally do not recommend the use of minoxidil after a hair transplant, unless it is being used in an area other than that which was transplanted (such as the crown). We feel that minoxidil has little added value for the average post-transplant patient. We do, however, encourage the continued use of finasteride to help retard further balding. Those who do not plan to use minoxidil after surgery should discontinue it immediately, so that any of its beneficial effects may be reversed by the time of surgery.

Patients are advised to avoid products that are used to thicken the hair or that stain the scalp for 3 days before the procedure, as these often take several days to completely wash out of the scalp. Their presence during surgery can decrease visibility and make placing more difficult. Hair systems should be removed before surgery and be replaced with, or converted to, a clip-on system. The front edge of the piece can be kept in place with a stiffening rod, obviating the need for tape or glue. However, any form of attachment at or near the frontal hairline risks dislodging grafts. Patients are encouraged to permanently discontinue the system after the procedure, but those who feel the need to use them until their hair grows in should wait at least 5 days post-op.

Systemic antibiotics are not indicated for clean surgical wounds in healthy patients, so their routine use in hair transplantation is not necessary; however, many doctors routinely use them.

Because there are no specific guidelines for antibiotic prophylaxis in hair transplantation for patients with a history of endocarditis or mitral valve prolapse, the decision to use them must be based on the individual patient's risk factors.⁴²

The other preoperative instructions are relatively straightforward and will depend to some degree on the preferences of the operating surgeon. Patients should be notified well in advance of the procedure date regarding the need to discontinue certain medications, to stop smoking and abstain from alcohol before surgery. Table 29.6 is a summary of the main preoperative instructions.

Table 29.6 Preoperative instructions
Continue medications currently prescribed by your physician, particularly those for high blood pressure. Those taking broad β -blockers (such as propranolol) should switch to select β -blockers, as the former may interact with epinephrine (adrenaline). This should be done under a doctor's supervision.
Discontinue topical minoxidil and avoid products used to thicken the hair or stain the scalp 3 days before the procedure.
In order to minimize bleeding, avoid the following before surgery: aspirin or other anti-inflammatory medications (1-2 weeks), vitamin B or vitamin E (1 week), or alcoholic beverages (3 days). Do not drink coffee or any other caffeinated beverages on the day of your procedure as these substances will increase your sensitivity to medications such as adrenaline.
Do not smoke tobacco products for at least 24 hours before your procedure and at least 1-2 weeks after. Smoking will slow down healing and heighten the chances of wound infection and scarring.
On the morning of your procedure take a bath or shower and wash your hair thoroughly using a surgical scrub (contains 3% chloroxylenol). If possible, leave your hair long in the back and on the sides to cover the sutures.
Wear clothes that do not need to be pulled on over your head; this will help keep your bandage in place and avoid any damage to your grafts immediately after surgery.
Eat breakfast the morning of your procedure. If you are scheduled for surgery in the afternoon, have a light lunch before you arrive.
Because you may be receiving medications during the procedure that can make you drowsy, you cannot drive home following the surgery. If you must drive or take public transportation, please let your doctor know ahead of time.

All patients undergoing hair transplantation should be treated with universal precautions. Although there is no consensus on the need to perform routine blood tests before the procedure, the following tests provide an additional level of safety:

- complete blood count
- hepatitis B surface antigen and antibody
- hepatitis C antibody
- HIV screen

One should obtain medical clearance for HIV-positive patients to make sure that they are immunocompetent enough to withstand a potential series of procedures. With the hepatitis antigen-positive patient, there is a concern about active disease and one should inform the patient's primary physician of a positive screening. Other tests, such as a platelet count, and prothrombin and thromboplastin times are performed only if warranted by the history and physical.

The surgical consent form should be given to patients to read at their leisure well in advance of the procedure and should be signed by the patient before any medications that can cause drowsiness or impair judgment have been given. The exact time of signing should be indicated. The major elements of the consent form are listed in Table 29.7.

Table 29.7 Elements of consent form for hair transplant
Nature of the procedure, with specific reference to its cosmetic nature Indications for surgery
Risks: reactions to anesthetics; allergic reactions; sterile folliculitis, infection; cyst formation at graft site; scarring in donor area; hair loss related to the procedure; hair texture changes; failure of transplanted hair to grow; numbness; paresthesia; temporary swelling or bruising
Benefits
Alternatives: doing nothing, change hair styles (e.g. lightening or keeping hair very short); medical therapy; another form of hair restoration surgery (e.g. mini-micrografting); wearing a hairpiece Results are not guaranteed
Results are not guaranteed
Consent for photograph as part of the medical record
Caution about driving under the influence of medication
Statement that the above are understood and all questions have been answered
Signature of the patient with date and time (before the administration of drugs), countersigned by the operating physician and witnessed by a staff member

Although not universally recommended, some doctors have the patient shower the morning of surgery using a chlorhexadine surgical scrub as shampoo. Though it does not sterilize the scalp, the chlorhexadine binds to the stratum corneum, decreasing transient and pathogenic microorganisms and resident skin flora. Caution is advised as it can be toxic to the middle ear and irritating to the eyes. In our practice, we advise patients to use a simple baby-shampoo. After showering, the patient should dress in comfortable clothes and wear a button-down shirt.

The Operating Room

Follicular unit transplantation is typically performed in an outpatient office setting. The guidelines of care for office surgical facilities delineated by the American Academy of Dermatology should be reviewed before setting up facilities.^{43,44} Though major complications occur only rarely, protocols should be in place to handle emergencies such as hypersensitivity

reactions or anaphylaxis, stroke, seizures, arrhythmias, acute myocardial infarction, and hypertensive crisis. At the very least, specific arrangements should be made with a local Emergency Medical Services (EMS) facility to take care of distressed patients.⁴⁵

The medical staff should be comfortable dealing with problems such as bleeding, syncope, petting episodes and anxiety reactions. The medical staff should be certified in cardiopulmonary resuscitation (CPR), and at least some in advanced cardiac life support (ACLS). The amount of in-office emergency equipment will depend on staff capability and training, and proximity to an EMS unit. Basic equipment includes portable oxygen, an automated defibrillator, intravenous setup, and an oral and nasopharyngeal airway.⁴⁵

Because of the scalp's abundant vascular supply and relative resistance to infection, it is common practice for doctors' technique to be aseptic, rather than sterile. And because the scalp is not usually shaved and prepped, strict sterile technique is not practical. However, at least until the donor area is sutured closed, a sterile environment should be maintained. This issue is discussed in greater detail by Sebben and Davis.^{46,47}

The single most important instrument for FUT is the dissecting stereomicroscope,^{8,48} which is available with either a binocular view or an LCD screen. There should be one microscope for each member of the surgical team performing dissection. Experienced teams generally require one staff member per 500 to 750 grafts (for graft dissection and placing), so that a typical 2000-graft procedure would necessitate three or four staff members in addition to the physician, though this will vary considerably among practices. Less-experienced teams may require more people. The staff's other responsibilities, as well as possible absenteeism, must also be taken into account.

Regular operating-room tables are generally inadequate. Rather, the tables must be contoured and provide some lumbar support when patients are sitting. They also need to be low to the ground, with the seat not more than 56 cm from the floor, so the staff can work comfortably around the head while the patient is seated.

Equally important are comfortable working areas for the staff. Dissecting tables with a bull nose, rather than squared edge, are easier on their arms, and adjustable seating saves them from bending over to look into the microscopes (Fig. 29.5). The long duration of the surgery makes meticulous attention to ergonomic issues all the more crucial.⁴⁹ Bright fluorescent ceiling lighting is preferable to surgical operating-room lights because it generates less heat. We use high-intensity surgical lights only for working on the patient's donor area, as they can be angled to illuminate the posterior scalp of the seated patient.⁵⁰

The Petri dishes containing grafts pending placement should be kept on a stable, wall-mounted surface to keep them from being inadvertently knocked over (Fig. 29.6). The typical operating room Mayo stand is not stable enough.⁵¹ Normal saline is the most frequently used holding solution for grafts, but we prefer the more physiologic lactated Ringer's. Limmer has shown a high survival rate for grafts kept in chilled saline for up to 8 hours.⁵² More exotic solutions with potential graft-sustaining properties, such as platelet rich plasma, have been developed, but they are not widely used.^{53,54,55}



Figure 29.5 Dissecting stereomicroscopes on cutting table.

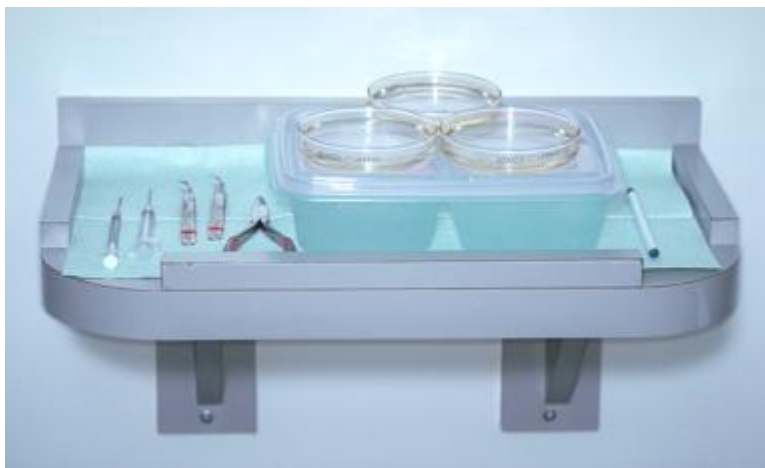


Figure 29.6 Wall-mounted placing stand for ice blocks, grafts, and instruments.⁵¹

Grafts should be refrigerated when they are outside the body. Approximately 1 hour before placement, we remove a portion of the grafts from refrigeration and put them into Petri dishes which are resting on ice packs that maintain the Ringer's solution at 59 °C. This is set on a wall-mounted stand conveniently located at the head of the operating chair.

It is preferable to have all the dissection performed in the operating room. In addition, a small refrigerator used solely for graft storage should be located in each room. Performing all the dissection and storing the grafts in the same room eliminates the risk of inadvertently placing grafts in the wrong patient. This is particularly important in a busy practice where more than one person is being operated on per day. The minimum room size for a physician and five staff members to work comfortably is approximately 15.5 square meters.

In advance of the procedure, it is helpful to set up complete surgical trays, including the local anesthetic mixtures pre-drawn into syringes. This serves a number of purposes: it makes it easier to keep track of how much anesthetic is used in each part of the procedure, it helps to ensure that safe dose limits are not exceeded⁵⁶ and it minimizes the risk of needle-stick injury, because the preparation takes place before the operating room begins to bustle.

Local anesthesia is administered with a ring-block consisting of lidocaine, bupivacaine, and epinephrine (adrenaline) buffered with sodium bicarbonate. Lidocaine, the major component, is used for its safety and quick onset. Bupivacaine is added to increase the duration of anesthesia, but in smaller quantities to limit its potential cardiac toxicity.⁵⁷ Epinephrine (adrenaline) increases the anesthetic duration while decreasing its toxicity and providing some hemostasis – although its vasoconstrictive action is relatively short-lived.

Sodium bicarbonate can be added to bring the acidic pH of the epinephrine (adrenaline)-containing solution closer to 7.4, lessening its sting.⁵⁸ This is particularly useful for injections at the hairline, where the scalp tends to be most sensitive. It is important to stress that the combined use of epinephrine (adrenaline) and broad β -blockers (e.g. propranolol) can result in potentially life-threatening reactions.⁵⁹

The anesthetic solution used for a typical 2000-graft session is listed in Table 29.8. The anesthetic is drawn into 13 5-ml syringes then fitted with 27-gauge needles. The tumescent mixture consists of three 10-ml syringes with 25-gauge needles. These quantities are preprinted on the operative report and circled as each syringe is used. If the patient requires additional anesthesia, the medication is drawn up and the additional doses are recorded.

Table 29.8 Anesthetic mixture		
Ring Block Component	Concentration	Amount in Mixture
Lidocaine	0.50%	≈ 60%
Bupivacaine	0.03%	≈ 40%
Sodium bicarbonate ^a	8.40%	01:10
Epinephrine	—	1:200000
Volume^b	Initial Quantity	Boost
Donor area	20 cc	10 cc
Recipient area	25 cc	10 cc
Tumescent mixture for donor harvest		
Component	Concentration	Amount in Mixture
Lidocaine	0.17%	≈ 100%
Epinephrine	—	1:600000
Volume^b	Quantity	
Donor area	30cc	
^a Used prn.		
^a Volume of mixture typically used in 2000-graft session.		

Techniques

On arriving at the office, patients sign the consent form which was given to them well in advance. The surgical plan is reviewed and any last-minute questions are answered. Next,

density is checked to confirm the measurement taken at the initial consultation. If the number differs or it appears that density varies in the donor area, multiple measurements are taken. Scalp laxity is also reassessed and recorded on the operative report.

Patients are given cotton surgical gowns to wear and their pictures are taken with a digital camera against a light-blue background. Next, the hairline and other important landmarks, such as the vertex transition point and crown swirl, are marked in Gentian violet and shown to the patient using two mirrors. If the patient approves of the hairline design, additional pictures are taken, this time of the marked scalp, usually from the front and aerial views. Three-quarter and close-up photos are sometimes taken to illustrate particular cosmetic issues. Occasionally pictures are taken during and after the operation for teaching purposes. All photos are kept as part of patients' permanent medical record.

Typically, for a procedure of 1,500 grafts or more, patients are pre-medicated with diazepam 5-15 mg P.O. and midazolam 2-4mg I.M. about 15 minutes prior to surgery. Dosing will depend upon the patient's body weight, age, sex, and past sedative/analgesic use. Long acting sedatives are not given to patients who will be driving themselves home from the procedure. Methylprednisolone 80 mg IM is given to reduce post-op swelling. Oral antibiotics are not routinely given, but are considered in patients with implanted prosthetic devices, valvular heart disease and other medical conditions requiring prophylaxis.

Donor harvest

In preparation for the donor harvest, patients are seated upright on the operating table. The hair in the donor area (that covering the donor strip plus the 0.5 cm beyond its perimeter) is then cut to 1–2 mm using electric clippers (the extra 0.5 cm is to facilitate suturing). The trimmings are thoroughly vacuumed away and the hair above the strip is held back with paper tape, fully exposing the donor area. A gauze headband is placed around the patient's head just below the clipped area and sterile drapes are taped to the headband.

In the first session, in a patient with average scalp laxity, we generally harvest a donor strip that is 1-1.3 cm wide. In calculating the length, we use the general rule of 90-100 follicular units per cm².⁵ For example, if a 1500 follicular unit graft hair transplant is planned for a Caucasian with average density, a 1 to 1.1-cm wide strip would need to be 18 cm in length (15 cm plus 3 cm for tapering). People of African and Asian descent characteristically have a lower follicular unit density and a strip of this dimension will yield a lower number of follicular unit grafts.⁷

Local anesthesia

A ring-block is established using an anesthetic mixture of lidocaine, bupivacaine and epinephrine (adrenaline) (Table 29.8). The anesthetic solution is injected into the subcutaneous fat layer approximately 1 cm below the lower portion of the clipped area and extending several centimeters past it on either side. In addition to buffering the anesthetic, already discussed, discomfort from the injections can be reduced by pressing a vibrating hand massager to the skin below the area insertion of the needle. The sting of the injection can also be reduced by a device that controls the flow of the flow of the anesthetic solution.⁶⁰

Approximately 0.75 ml of anesthesia is used per centimeter of donor area, so that a 25-cm-long incision would require slightly less than 20 ml of anesthetic solution. It is important to avoid injecting into the muscle, as epinephrine (adrenaline) will cause vasodilatation, due to the action on β_2 -receptors, quickly dissipating the local effects of the anesthetic and increasing its toxicity.⁶¹

The ring-block takes approximately 15 minutes to induce anesthesia. As soon as the donor-area skin becomes numb, tumescent anesthesia is administered by injecting larger quantities of a more dilute lidocaine epinephrine (adrenaline) solution into the mid-fat to make the area firm. The tumescence serves six purposes: 1) to widen the distance from the follicles residing in the upper fat to the nerves and larger blood vessels lying just above the fascia, 2) to increase the rigidity of the donor area, 3) to decrease follicular transection, 4) to decrease bleeding, 5) to produce more uniform anesthesia, and 6) to reduce the amount of anesthetic required.

Tumescence can be achieved using a dilute solution of epinephrine (adrenaline) and lidocaine (Table 29.8) in somewhat higher concentrations than used in liposuction surgery, as this will provide additional anesthesia that is particularly important in repeat procedures, or for patients with significant donor scarring.⁶² In situations where there is prior surgery, direct cutaneous innervation to the donor area from the occipital branches can be blocked by scar tissue, so that innervation to the donor area arrives in a rostral–caudal direction, rendering inferiorly placed ring-block anesthesia ineffective. At times, when there is an excessive amount of donor scarring, the anesthetic mixture must be injected above or directly into the scarred area to make it numb.

Strip excision

The ideal position for the donor incision is in the mid-portion of the permanent zone. This generally lies at the level of the external occipital protuberance. The donor strip can be excised using two parallel blades set 1.0 to 1.5 cm apart or the strip can be harvested using a single blade. When using the parallel blade technique, a convenient harvesting device is the Rassman handle that is loaded with two #10 blades. The handle holds the blades at an angle of 30°, making them parallel to the emergent hair. The handle should hold the blades in a pre-angled position to allow both blades to lie flush with the skin surface when aligned with the hair follicles; otherwise, the surgeon may cut too superficially with the upper blade and too deep into the scalp with the lower (Fig. 29.7). Once the main parallel portion of the incision is complete, a single scalpel blade is used to taper the ends into an ellipse. In general, the length of each tapered end should measure at least one and half times the width of the strip, so that the ends lie flat (Fig. 29.8).



Figure 29.7 Donor-strip harvesting using two parallel blades on a Rassman handle pre-angled at 30° to allow both blades to lie flush against the scalp.



Figure 29.8 Dissection at the tapered corner of the strip in a mid-subcutaneous plane, just below the follicles.

Alternatively, the entire excision can be performed with a long free-hand ellipse after the skin is marked.⁸ This allows the angle of the blade to be adjusted as each edge is cut. On the other hand, this technique makes it harder to keep the width uniform, which is necessary for predictable graft yields. Cutting the second edge also becomes more difficult because the rigidity of the tumescence is lost. The first incision renders the second wound edge mobile and there is more distortion of the wound edge and hair follicles due to the relative elasticity of the dermis compared to the epidermis and fat.

Once the strip is removed it is immediately placed into a chilled Ringer's lactate solution.

Donor closure

The most common types of closures include metal staples, a running non-absorbable suture, or a running absorbable suture.⁶³⁻⁶⁵ If staples are used to close the wound edges, it is important to be certain that the wound edges are flush before the staples are applied. The edges can be approximated by grasping the lower edge with a skin hook and using rat-toothed forceps to hold and slightly evert the upper edge (this requires the help of an assistant). The staples should then be placed approximately 0.6 cm apart (Fig. 29.9). Staples have virtually no tissue reactivity, but they make flush apposition of wound edges difficult and can be uncomfortable in the post-op period and sometimes painful to remove.⁶³ In the author's practice, we routinely remove alternating staples at 9-10 days and the remainder at 18-20 days post-op. We feel that leaving in staples for this longer period of time leads to a finer donor scar, particularly in patients with tighter than average closures. This opinion has yet to be confirmed in controlled studies.

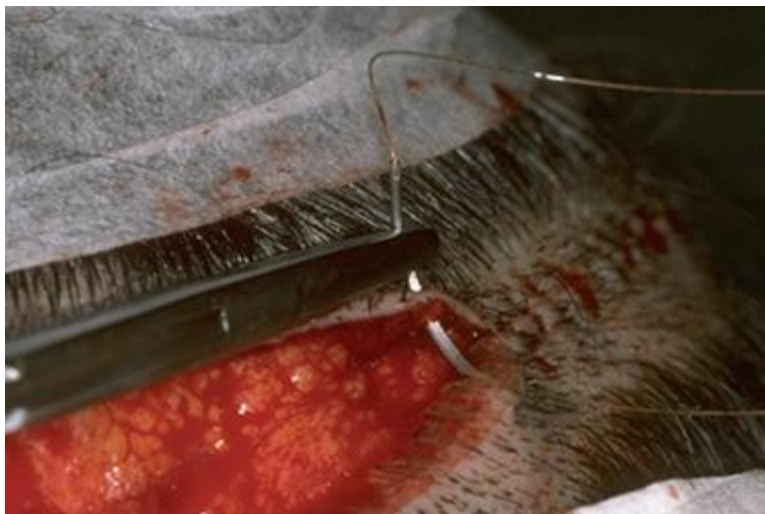


Figure 29.9 Suturing the donor area using absorbable sutures. Note the needle track is parallel and very close to the wound edges.⁶³

Non-absorbable sutures, of either nylon or polypropylene, are usually placed using a single, running stitch and are removed in 10 to 14 days. A problem is that if they are placed too far from the wound edge they entrap hair follicles and this may result in the destruction of donor hair if there is significant post-op edema. If placed too close to the wound edge they can become buried and difficult to remove. These two problems are avoided with the use of staples and speak in part to why staples are used by many hair transplant surgeons in spite of the preference that patients have for the more comfortable sutured closure.

Absorbable sutures, should be placed close to the wound edge. This will tend to minimize the amount of suture material used, decrease the entrapment of follicles and avoid strangulation if there is significant post-operative edema. To further limit damage to follicles, the sutures should be placed approximately 4.5 mm apart and the suture should be advanced on the surface, rather than under the skin, as this will minimize the amount of suture in contact with the follicles (Fig. 29.10). With this technique, the needle should be passed through the full thickness of the dermis and exit the wound edge, just below the level of the bulbs, without incorporating any significant

amount of subcutaneous tissue. The needle track must be kept parallel to, and within 1.5 mm of the wound edge (Fig. 29.11). Particular attention should be paid to placing the needle parallel to the upper wound edge where the angle is very acute. Basic guidelines for using poliglecaprone 25 sutures can be found in Table 29.9.

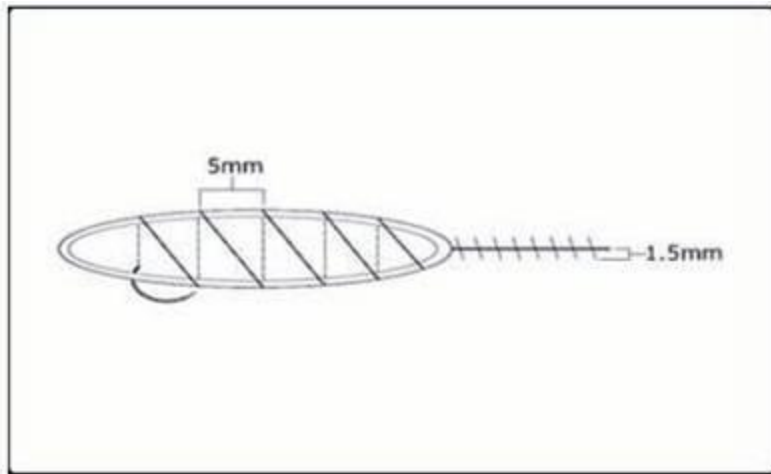


Figure 29.10 Schematic of the suturing technique recommended for sewing the donor scalp. The spacing between loops is 4.5 mm.⁶³



Figure 29.11 Stapling technique illustrating very controlled apposition of wound edges, using skin hooks for the lower edge, and forceps to evert the upper.⁶³

Table 29.9 Guidelines for Using Poliglecaprone 25 Sutures
Plan width of donor strip so that there is little or no tension on closure
Use tumescent anesthesia to harvest donor strip in mid-fat
Use sutures no heavier than 5-0
Use a simple running stitch, advancing each loop on the skin surface
Keep needle parallel to and within 1.5 mm of wound edge
Incorporate epidermis and dermis only
Use 4.5 mm spacing between loops

Graft dissection

Once the donor strip is removed, it is immediately placed into lactated Ringer's, or similar holding solution, and passed to the surgical team leader stationed at a stereomicroscope (Fig. 29.12). The strip dimensions are measured and recorded. The next step is performed under strict stereomicroscopic control in a process called 'slivering.'^{27,39}



Figure 29.12 Close-up view of a donor strip showing the natural hair groupings at the surface, the slightly curved path of the hair through the dermis, and the random distribution of bulbs in the fat. It is this unique anatomy that makes stereomicroscopic dissection essential in preventing follicular transection.

In one method of slivering, the donor strip is placed on its side, on a wooden tongue-depressor blade, soaked in Ringer's, with the hair pointing away from the dissector and the convex surface of the strip facing upward. For a right-handed person performing dissection, the left end of the strip is held with rat-toothed forceps in the dissector's left hand. An assistant applies tension to the strip, holding it a few centimeters away from the area being cut with rat-toothed forceps held in the right hand. Using a Persona #10 blade on a #3 blade handle held in the right hand, the dissector begins to cut the strip into 2–2.5 mm wide sections, by guiding the blade between follicular units using a one-directional fillet-like movement from the epidermal side to the subcutaneous surface. A back-and-forth sawing motion should be avoided as it will increase the

risk of transection (Figs 29.13, 29.14). The pieces generated are then passed along to the other members of the surgical team who complete the dissection. In the last step, the individual pieces are placed on their sides, stabilized with straight jeweler's forceps, and then dissected under the stereomicroscope with a scalpel into individual follicular units of one to four hairs (Figs 29.15, 29.16).



Figure 29.13 'Slivering' the donor strip into sections.

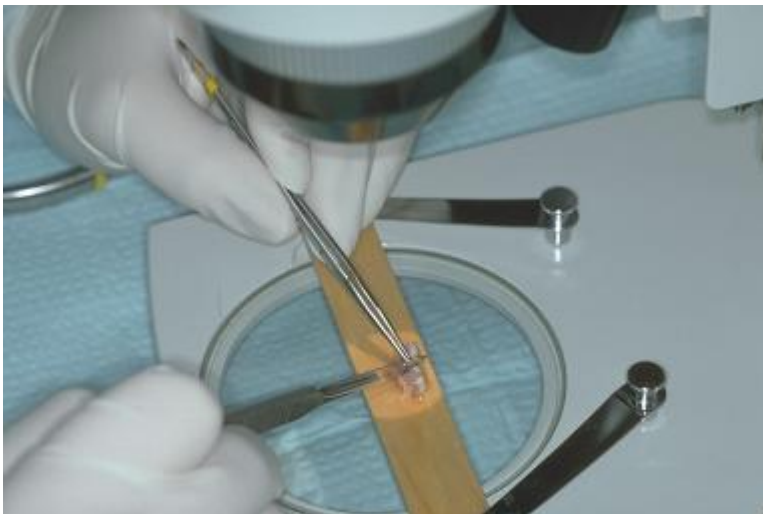


Figure 29.14 Stereomicroscopic dissection of one section into smaller pieces.



Figure 29.15 The final step of the dissection: generating individual follicular units.

Next, the units are sorted according to the number of hairs they contain into separate Petri dishes held on ice blocks and filled with Ringer's lactate (Fig. 29.17). When a substantial number of grafts (several hundred) have been dissected, they are put into secure plastic specimen containers and refrigerated.



Figure 29.16 Dissected one-hair, two-hair, three-hair, and four-hair follicular units.



Figure 29.17 Petri dishes with one-hair to four-hair follicular units immersed in Ringer's lactate.

An alternate slivering method involves dissecting the donor strip into slivers as wide as one follicular unit, approximately 1 mm. The units are then isolated from the very thin strip. The key to either method is that every step is performed under stereomicroscopic control, which keeps the units intact and avoids follicular transection. It is also vital that all the pieces of the strip remain in chilled Ringer's lactate except when they are being cut. While under the microscope they should be kept well-hydrated using 10-ml syringes containing Ringer's lactate that is kept at each cutting station.

Follicular Unit Extraction

Follicular Unit Extraction (FUE)¹⁰ is a conceptually simple procedure, where individual follicular units are harvested through small circular incisions created by a trephine or similar instrument. The punch cuts into the reticular dermis and the remainder of the follicular unit is literally extracted from the scalp. The advantage of this procedure is that it obviates the need for a linear incision. Difficulties lie in the intrinsic variability of each patient with respect to the ease of extraction, the increased risk of follicular transection compared to other techniques, the inability to harvest all the hair from the mid-portion of the donor area, and the organizational limitations of not being able to have multiple persons (more than two) working in parallel, as can be done with the microscopic dissection of a harvested strip.

FUE is appropriate for patients who have limited hair loss and who want to wear their hair so short that a linear donor scar might become visible. The procedure is also useful in patients who have healed poorly, or who have very tight scalps. An important application of this technique is to camouflage a linear donor scar from prior hair transplant procedures.¹⁰

FUE's main limitation is that it is less efficient in harvesting hair than follicular unit transplantation performed with strip harvesting. In FUT, all the hair from the optimal (central) part of the donor region can be removed and transplanted, and the resulting defect is sewn closed. In contrast, the FUE defects remain open to heal by second intention with significant amounts of intervening hair left behind and, therefore, a much larger region of the scalp must be

accessed to harvest the necessary amount of donor hair. The greater area required, greatly increases the risk of harvesting hair from non-permanent areas of the scalp. Figure 29.18 shows the donor area immediately after the extraction of 600 follicular units and the same area at 6 and 13 days post-op.



Figure 29.18 Donor area in follicular unit extraction. (A) Immediately after harvesting 600 grafts. (B) Six days post-op. (C) 13 days post-op.

With FUE, the second intention healing causes microscopic fibrotic changes in the donor area, distorting adjacent follicular units and making subsequent sessions more difficult. As mentioned, transection during the harvest is greater than stereo-microscopically controlled FUT. Taken together, these factors limit the total amount of hair that can be accessed through FUE, rendering it a less robust procedure than Follicular Unit Transplantation. In a procedure where a finite donor supply is the main constraint, the inefficient use of donor tissue poses a significant problem. Table 29.10 summarizes the pros and cons of this follicular unit extraction.

Pros	Cons
No linear scar	Requires multiple sessions to equal the size of a single follicular unit transplantation procedure
Useful when the patient wants (or needs) to wear the hair styled back from the face and with the sides very short	Takes longer to perform and is more expensive than follicular unit transplantation
Provides an alternative when the scalp is too tight for a primary closure or when the patient heals with widened scars	Risks the eventual loss of transplanted grafts by harvesting from outside the 'sweet spot' of permanent hair in the donor zone
Ideal for camouflaging donor scars that cannot be excised	After large numbers are harvested, fine stipple scars may become visible due to thinning of donor area
Makes it possible to harvest non-scalp hair (e.g. beard or body hair)	Scarring and distortion of follicles may make farther harvesting by follicular unit extraction difficult
	Maximum possible yield lower than with follicular unit transplantation

The ability to perform FUE with minimal transection varies significantly among patients. Because of this, a test measuring the ease of extraction (the FOX Test)¹⁰ should be performed prior to recommending the procedure. Patients undergoing follicular unit transplantation should also be tested for follicular unit extraction at the time of surgery, in case the latter procedure is considered at a future date.

A significant advance in the technique of follicular unit extraction is to first score the top of the follicular unit with a sharp punch and then use a blunt instrument to separate the follicular unit from the surrounding tissue, thereby minimizing follicular transection. The main limitations of this technique are the increased incidence of buried grafts and increased surgical time in an already very laborious process.^{66,67} It is encouraging that follicular unit extraction techniques and instrumentation continue to improve and some progress had been made towards automating the extraction procedure using robotics.⁶⁸

Recipient sites

The way recipient sites are prepared determines the most critical aesthetic aspects of the transplant: the angle at which the new hair grows, its distribution, its density, and how natural the hair will look as it emerges from the scalp. A thorough discussion of site creation is beyond the scope of this chapter, but the four basic elements will be briefly reviewed. These are recipient site size, hair direction, site density, and graft distribution, and they are covered in greater detail in “Aesthetics of Follicular Transplantation.”⁷

Recipient site sizes and orientation

The sites should provide a snug fit for the graft: just large enough to accommodate a follicular unit, but not so small that insertion is difficult or traumatic.¹ Numerous instruments have been developed to create recipient sites. The main aspect to consider in choosing one is its diameter.⁶⁹ Although there are many viable options, the following are used by many practitioners:

- 21-gauge hypodermic needle for 1-hair follicular unit grafts
- 20-gauge for thin 2-hair follicular unit grafts
- 19-gauge needle for thick 2-hair and all 3- and 4-hair follicular unit grafts
- 18-gauge for 3-hair grafts in African races with very kinky hair

These needles can be fitted on 1- or 3-ml syringes for ease of use, obviating expensive handles. Some physicians advocate smaller recipient sites, but this may require dividing four-hair follicular units—a technique that this author does not recommend.³⁹ For very fine hair, or in special situations like transplanting eyebrows, a 22-gauge (or finer) needle can be used.

As an alternate to using hypodermic needles, some physicians use a blade-breaker that enables a straight blade to be custom cut into chisel shaped cutting instruments of various sizes. It is this author’s opinion that hypodermic needles, which spread apart the skin as the wound is created, tend to cause less tissue injury than chisel-shaped blades, which cut through the skin.

Most instruments used to make sites create tiny slits as they cut through the skin and these slits can have either a coronal (horizontal) or sagittal (vertical) orientation. The advantages of coronal incisions are: a more natural appearance, since the original orientation of most follicular units seems to be coronal; a fuller look; and less tendency of the growing hair to elevate in the vertical plane. The advantages of a sagittal orientation are: greater visibility of sites; ease of graft placement; less trauma to existing hair; and less lateral (radial) splay of hair. Because coronal incisions provide better coverage and a slightly more natural look, they are currently used by the majority of hair transplant surgeons.

Hair direction

Hair should be placed into the scalp at the angle it originally grew, not in the direction that it is to be groomed. However, there are some exceptions. Whorls at the frontal hairline should not be mimicked with a hair transplant, nor should hair at the hairline be planted in a radial direction, even if some of the original hair grew that way. In general, hair anterior to the vertex transition

point should point forward, with the angle becoming more acute as it reaches the anterior hairline – where it is essentially horizontal to the ground – regardless of the slope of the forehead. By planting the hair in the frontal hairline in forward direction (rather than radially) the transplanted hair will act more as a unit and hold together when it is groomed. (Fig. 29.19).⁷ At the temples, the forward pointing hair makes an abrupt transition to a downward direction. Hair at the temples should be transplanted at a very acute angle, i.e. lying as close to the surface of the skin as possible.

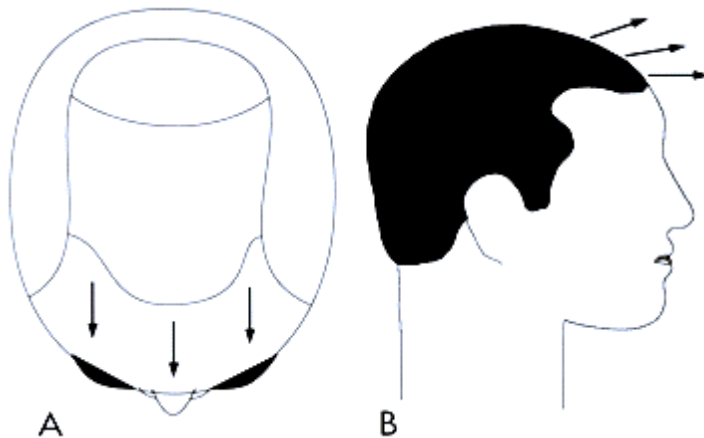


Figure 29.19 Normal hair direction. (A) Top view and (B) side view.⁷

Recipient site density

The average non-balding scalp has 90-100 follicular units per cm². Approximately 50% may be lost before there is any noticeable thinning. It would be wasteful for more than 50% to be replaced, especially since hair transplants are always performed in the face of diminished total hair volume. We generally recommend transplanting up to 25-30 follicular units per cm² into the frontal area of a balding scalp in the first session. If the larger three-hair and four-hair units are placed in select areas, more than 25% of the initial density can be achieved in one pass. The ideal transplant density can be achieved in many patients with two procedures.⁷

Some physicians advocate a ‘one-pass’ procedure to achieve the final density.³⁹ Although this may be appropriate for some patients, the increased incidence of graft popping and desiccation, insertion injury, and possible vascular compromise may lead to poor growth. For very bald patients, very dense packing does not allow coverage of an entire bald area.

Transplants of more than 3,000 follicular unit grafts may necessitate that the grafts be out of the body so long that their survival may be diminished. In addition, the large donor strip required to harvest graft numbers of this magnitude increases the incidence of a widened donor scar. Furthermore, for procedures of 3,000 or more to be practical, the recipient area should be shaved, a condition unacceptable to many patients. It is this author’s view that covering the entire bald area and then increasing density in a subsequent session is a better goal for the majority of patients.

Distribution

For simplicity, the area of the scalp subject to androgenetic change can be divided into three regions: the frontal region that includes the frontal hairline; the top or mid-scalp; and the vertex or crown. The vertex transition point separates the top of the scalp from the crown (Fig. 29.20).³⁸ As most social interaction takes place face to face, and people generally view themselves from the front, the overall impact of the transplant is essentially defined by the position of the frontal hairline and density of the frontal region of the scalp.

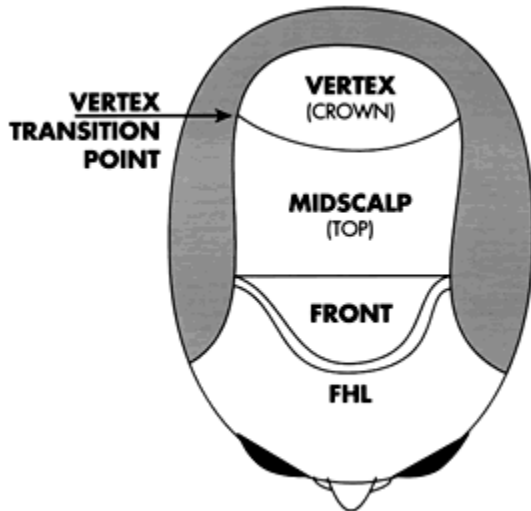


Figure 29.20 Regions of the scalp.

For most patients who are moderately or extensively bald, or for those who are destined to be, the limitations of the donor supply make restoring the entire bald scalp to ideal density impossible. Consequently, creating the greatest density in the frontal forelock area of the scalp produces the best cosmetic result. This 'forward weighting' can be accomplished by putting recipient sites closer together, placing larger follicular units (i.e. those with three and four hairs) in those sites, or by a combination of both.

In general, recipient site density should be the highest in the front part of the scalp and gradually tapered toward the crown. In contrast, the largest follicular units should be placed in the forelock region of the scalp. This overlapping distribution of sites and follicular unit grafts may be visualized in Figure 29.21. It is explained in greater detail in 'The Aesthetics of Follicular Transplantation.'⁷ The pattern serves two functions: 1) it creates a natural central-forelock density without the need for spacing sites closer towards the mid-scalp, where the blood supply can most easily be compromised, and 2) it insures the most natural look by placing the larger follicular units in a forward-central position, but away from the frontal hairline. Density in the forelock area brings about a 'patterned look' and avoids the diffuse thinning that often results when small grafts are evenly distributed over the scalp.



Figure 29.21 Schematic of ‘forward weighting,’ achieved by close placement of recipient sites (shaded dark grey), and central density, accomplished by placing larger follicular units in the forelock area (dark brown oval).

Practically every patient has enough donor hair to provide at least light coverage extending to the vertex transition point. This is a natural stopping point because, even if the crown continues to enlarge, transplants performed to this point will continue to look natural without additional surgery. Transplants should be extended past the vertex transition point into the crown only when there is adequate hair to create a swirl and follow the hair loss if the balding progresses. The indications for transplanting the crown are discussed in detail in “Follicular Transplantation: patient evaluation and surgical planning.”⁶

The distribution of grafts in the first transplant session should, in general, be symmetric. However, once the first transplant has had a chance to grow and the patient is willing to commit to styling his/her hair in a specific pattern, ‘side weighting’ may be considered. This is accomplished either by placing a greater proportion of the grafts on the parted side of the scalp, using larger follicular units on that side, or doing both. It results in increased fullness when one’s hair is combed to the side. In situations where there is a great imbalance in the supply/demand ratio, such as after scalp reductions or old plug procedures, a more exaggerated form of ‘side weighting’ should be considered. In this case the follicular unit grafts are concentrated on the front and part side of the scalp, but widely scattered in the top and back (Fig. 29.22).⁷

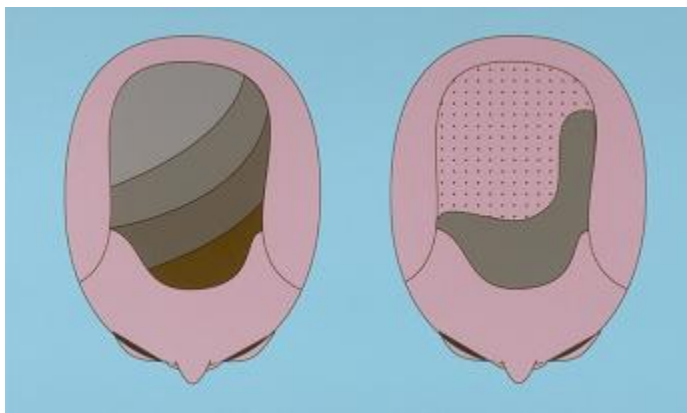


Figure 29.22 Left shows schematic view of left-to-right 'side weighting.' Right shows extreme side weighting employed when the donor supply is severely diminished.

When the follicular units are completely dissected and grouped in Petri dishes, they are placed at the head of the operating table on ice blocks. Graft placement, carried out in a front-to-back pattern, begins at the frontal hairline starting with one-hair follicular units. In a typical transplant, approximately 250 one-hair units are used for the leading edge of the frontal hairline, immediately followed by two-hair grafts. Three-hair to four-hair units are concentrated in the forelock area, while toward the back and sides the pattern is reversed so that the three-hair and four-hair units are always central to the one-hair and two-hair units.

Approximately 20–30 grafts are transferred at one time from the dish to the index finger, held on the finger in a droplet of Ringer's solution. The grafts are inserted using curved jeweler's forceps. (Fig. 29.23) A small band-aid should be worn on the finger under the area the grafts will sit to decrease the incidence of a stick injury from the forceps. Each graft should be carefully grasped by the fat at its bottom or at its edge, not at the germinative center. In the two-handed technique, gauze—held between the free fingers of the hand holding the grafts—is used to keep the partially inserted grafts in place as the forceps are repositioned higher along the length of the graft, to facilitate the last phase of insertion. This is a particular useful technique when there is popping (the tendency of grafts to lift up as they are being placed.)



Figure 29.23 Staff using loop magnification and curved jeweler's forceps to place follicular unit grafts into pre-made recipient sites.

Most physicians make the majority of recipient sites prior to graft placement. This allows the doctor to concentrate on the angle and distribution of sites without having to worry about actual graft placement. It also allows the team to easily clean the recipient area without having to be concerned about dislodging grafts. In addition, it gives time for some coagulation to occur and for the sites to become more "sticky" as this holds the grafts in place and reduces the incidence of popping.

In an alternative method, termed ‘stick and place’, the grafts are inserted at the same time that the recipient sites are made.³⁹ In this technique, the needle that is used to make the recipient hole can serve as a ‘shoehorn’ to help guide the graft into the site. The advantage of this method is that it eliminates the possibility that sites may be left unfilled or that two grafts may be placed into one site (piggybacking). It also moves the surgery along faster, since the staff does not need to wait for the surgeon to make the sites and they do not need to search for the sites that are empty.

On the other hand, there is more bleeding when sites are made concomitantly with graft placement (decreasing visibility) and there is an increased risk of the grafts popping and then drying out on the surface of the scalp. In addition, the staff must make judgments regarding the angling and distribution of the sites they are creating at a time when they must also be focusing on the technical aspects of graft insertion. This leaves many of the aesthetic decisions in the hands of the technicians, who are actively engaged in stick and place, rather than with the physician who could take a more strategic view of the procedure.

Patient disposition

After graft insertion is complete, the scalp is cleaned with distilled water, bacitracin zinc ointment is applied to the sutures and a head-band type pressure dressing is placed over the donor area. The transplanted area is covered with a surgeon’s cap. Prior to leaving, patients are given verbal and printed postoperative instructions, a neck pillow and medications for sleep and pain (Table 29.11). Patients leave the office wearing a bandana covering the cap and the headband. It is worth stressing that patients given sedatives or pain medication should not drive after surgery.

Class	Medication	Use
Topical antibacterials	Hibiclens	Preoperative
Systemic antibiotics	Dicloxacillin, cephalosporin	Intraoperative prophylaxis*
Sedatives/hypnotics	Diazepam, aprazolam ⁹⁸	Intraoperative sedation
Corticosteroids	Betamethasone	Postoperative swelling
Topical antibiotics	Bacitracin ointment	Postoperative care, donor area
Opiates	Hydrocodone	PRN, postoperative pain
Antihiccup agents	Chlorpromazine	PRN, hiccups
Analgesics	Nitrous oxide ⁶¹	PRN during injections [†]
Wound healing promoters	Copper-peptide	Postoperative healing [†]
Hair growth promoters	Finasteride, minoxidil	Pre and postoperative*
*See text.		
[†] Not used in our practice.		

OPTIMIZING OUTCOMES

Transition zones

A soft natural hairline is the hallmark of the ideal hair transplant. However, careful observation of a normal frontal hairline reveals that there is no actual 'line,' but rather an irregular, slightly asymmetric 'transition zone' or gradation of follicular units of increasing size and density. This is what should be replicated in the transplant (Fig. 29.24A–D). Transition zones are not limited to the frontal hairline; they must be recreated wherever the edge of the transplanted area is visible. In Figure 29.24C, one notes that transition zones have been created at the frontal hairline, sides and vertex transition point leading to the crown.

Although there are an infinite number of ways to create a transition zone at the frontal hairline, the placement of approximately 75–100 single follicular units in front of a band of staggered single units, one to three rows deep, will usually achieve the appropriate softness. This should be followed by two-, three- and eventually four-hair units, but the larger units should be more centrally located (Fig. 29.24B). With finer hair, larger units may be placed closer to the hairline. It is important to limit the depth of the single hair units, or the zone will appear too thin.

Visibility

Pre-making all the recipient sites before graft insertion helps to control bleeding and limits the need for epinephrine (adrenaline) (Fig. 29.24C). It initiates the 'extrinsic pathway' so that coagulation can begin before the grafts are introduced and allows for easy cleaning of the recipient area of any blood or coagulum, without the risk of dislodging grafts. It also makes it possible to place sites close together without a concern for grafts in adjacent sites from popping. Most importantly, it provides maximum visibility in the placing phase of the procedure.⁵⁰

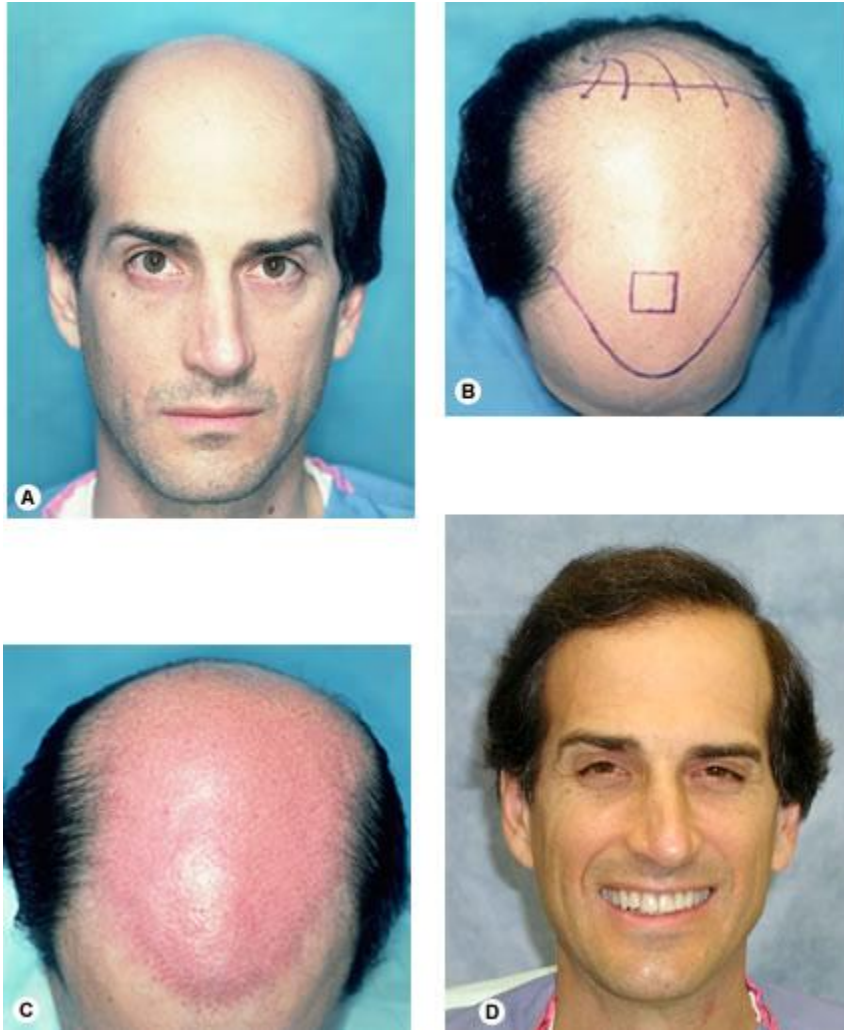


Figure 29.24 (A) Before and (D) 1 year after two sessions totaling 4295 follicular unit grafts. (B) Preoperative markings of the planned procedure. (C) First session just before placement, showing the distribution of 2355 recipient sites.

If the initial sites are scattered, they tend to cause the cutaneous vasculature of the scalp to ‘clamp down’, increasing visibility for subsequent passes and allowing the skipped areas to be filled in. When pre-making sites, the logistics of matching the number of sites to the number of grafts can be solved by making ‘projections’ of the anticipated number of grafts while the dissection phase is still in progress.

Recipient site influences

Some hair characteristics, such as waviness, are governed by the recipient site. When individual follicular units are placed into very small wounds, there is little fibrotic reaction to the grafts and the recipient area can exert its influences on hair growth. In Figure 29.25, note the delicate wave produced by the recipient scalp. It sometimes takes a year or more for the hair character to return to normal.



Figure 29.25 Norwood class VI, with very fine, blonde hair, and a donor density of 2.5 hairs per mm². (A) Before and (B) 1 year after two sessions of 2678 and 1836 follicular unit grafts.

Hair direction

When the hair is planted at the frontal hairline in its natural forward-pointing direction, combing it back causes it to bow, giving it the appearance of fullness. A common mistake is to point the hair in the direction that the patient plans to comb their hair; this makes the hair lie flat and bodiless when groomed. On the other hand, as the frontal hairline approaches the temples, the hair direction should be abruptly changed, so that it is angled downward and lying very flat on the scalp. Perfect integration of the frontal hairline with hair on the sides and temples is essential if the transplant is to look completely natural (Fig. 29.26).

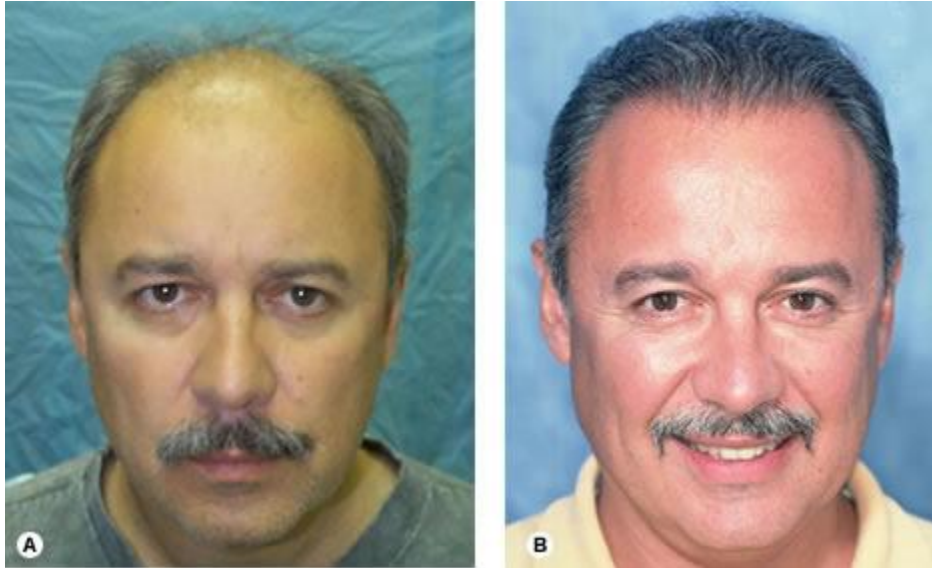


Figure 29.26 Early Norwood class VI. Before and (B) 1 year after one session of 2520 follicular unit grafts.

Hairline placement

If it is to look natural throughout the patient's life, the transplanted hairline must simulate that of a mature adult. In the adolescent male, the frontal hairline touches the top crease of the brow when the eyebrows are raised (that is, the hair begins at the upper border of the frontalis muscle). In a mature adult, the mid-portion of the frontal hairline rests approximately one finger-breadth (1.5–2 cm) above the brow crease. Although the degree of temple recession can vary dramatically in the normal adult hairline, the mid-part of the frontal hairline is crucial in framing the face.^{1,7} Significant bitemporal recession was built into the restoration of the patient in Figure 29.27, yet the forward-placed mid-portion sets his facial features in the correct proportions.

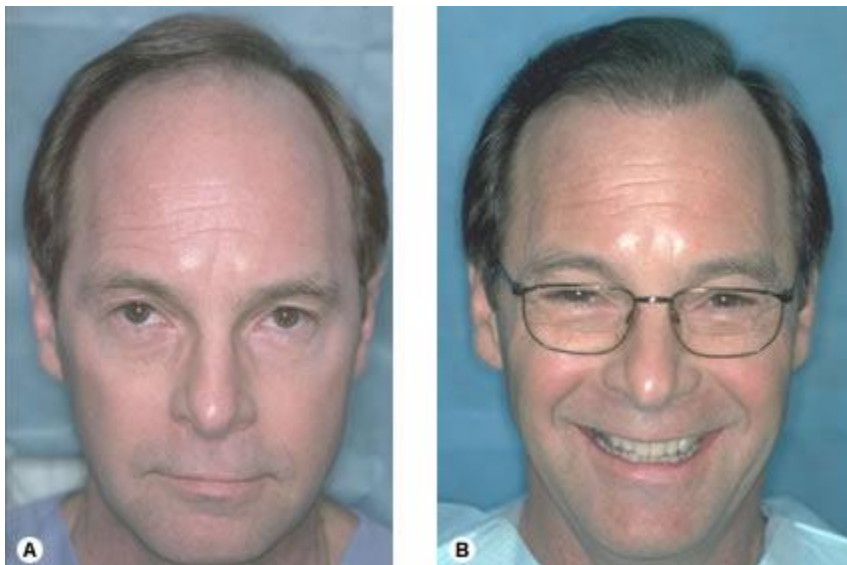


Figure 29.27 Early Norwood class V. Before and (B) 1 year following two sessions of 2105 and 1665 follicular unit grafts.

Forelock integration

A persistent frontal forelock poses an interesting dilemma. When hair is transplanted around the forelock, the patient may be left with a gap in his frontal hairline if the forelock eventually disappears. On the other hand, it is wasteful to add hair to a forelock with adequate density that may persist for years. The solution is to check for miniaturization with a densitometer. The patient in Figure 29.28 had miniaturization in his forelock greater than 50%, suggesting that it would not be stable over time. Two sessions were used to integrate transplanted hair into the resident terminal hair of the forelock, producing a central density that will be permanent.



Figure 29.28 Norwood class V. (A) Before procedure. (B) Integrating hair with a persistent frontal forelock (1 year following two sessions of 2133 and 1171 follicular unit grafts).

Optimizing density

One of the advantages of FUT over micro-grafting techniques is follicular unit sorting, the ability to sort the three-hair and four-hair units and place them in the forelock area, creating a density significantly greater than if the grafts were evenly distributed. This is possible because the compactness of the follicular unit allows four-hair grafts to be placed into very small sites.^{5,7} In certain instances, follicular units may contain more than four hairs, and adjacent follicular units are occasionally so close together that they can fit into a small recipient site.^{70,71} However, as long as these 'grouped grafts' are placed in a central location and the recipient wounds are kept small, this technique can add density without risking an unnatural look. It is important to keep in mind that ultimate density depends on the absolute number of hairs moved rather than the size of the grafts.⁷² Using grafts larger than follicular units always risks that the transplant may look unnatural. The patient in Figure 29.29 achieved a very dramatic frontal density in two sessions using the technique of follicular unit sorting.



Figure 29.29 A 46-year-old early Norwood class IVa with wavy medium-coarse hair. (A) Before transplant. (B) After two sessions of 2085 and 1438 follicular unit grafts.

Creating camouflage

A sizable part of many practices involves repairing transplants performed with older techniques. FUT is ideal for this because of the small size of follicular units and high hair count.^{73,74} Often fixing a ‘pluggy’ look requires graft excision with re-implantation, before the actual transplant. Although, the patient in Figure 29.30 had a row of plugs set far enough back from his frontal hairline that camouflage with follicular units alone was sufficient for the repair, in general, it is better in the long-run to excise and redistribute questionable grafts as individual follicular units than to attempt to camouflage them.

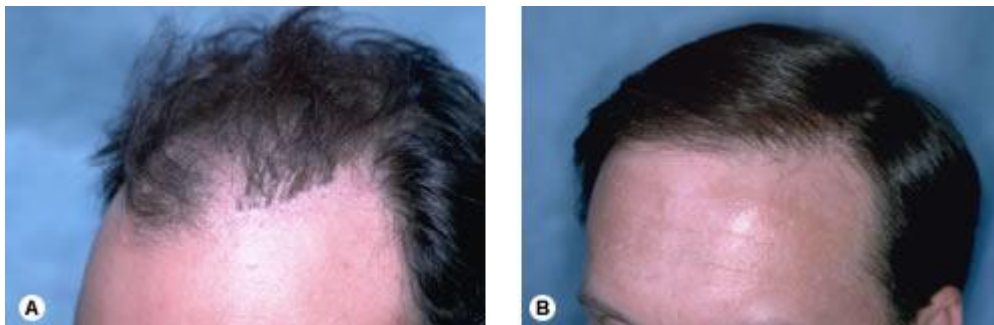


Figure 29.30 (A) Before transplant, showing large grafts forming a literal wall of plugs. (B) Camouflage from one session of 1818 follicular grafts.

Eyebrow Transplantation

Eyebrows are perhaps more important to one’s appearance than scalp hair, as eyebrows are in a more central position on the face and serve to frame the eyes. Unlike the loss of scalp hair, the loss of one’s eyebrows is not viewed as a natural process and is, therefore, not cosmetically acceptable. Because eyebrows have their own unique attributes, eyebrow transplants require techniques different from hair transplants on the scalp: 1) eyebrow hairs grow as individual strands, therefore multi-hair follicular units should be divided into 1-hair micrografts before planting, 2) eyebrow hair must be planted at a very acute angle so that the hair grows flat to the skin’s surface, 3) subtle angle changes are required to recreate the fan-like splay of hair at the medial aspect of the brow, 4) the hair must be planted in a crisscross growth pattern that causes

the hairs in the middle of the eyebrows to converge and form a subtle natural elevation, and 5) since the hair cycle of scalp hair (3-7 years) is longer than eyebrow hair (4 months) the eyebrow hair (after a transplant from the scalp) must be periodically trimmed.⁷⁵

Eyelash Transplants

The donor hair used in an eyelash restoration must match, as closely as possible, the original eyelash. Hair suited for this purpose includes the fine short hairs at the nape of the neck, hair from behind the ears or the eyebrows. The incision made into the lid margin should be made with a curved instrument and the hair should be rotated to follow the curvature of the original lashes.⁷⁶ In a very elegant procedure, scalp hairs are threaded onto a crescent needle and then introduced into the eyelid through an incision made on the inside of the lid margin. The hairs were then cut to the appropriate length.⁷⁷

Long-Hair Transplants

Transplanting long hairs, instead of the traditional 1-2mm clipped hairs, allows patient to temporarily see the actual results immediately upon completing the procedure. In this technique, the donor hair is left long (up to 25cm) throughout the harvesting, dissecting, and placing steps. As with traditional hair transplants, the hair is shed about 4 weeks after surgery. Although the procedure gives the patient an early view of the results and eliminates the problems of piggy-backing and buried grafts, it is technically more laborious and requires more meticulous post-op care.^{78,79}

Trichophytic Donor Closure

A Trichophytic closure is a way of improving the appearance of the linear donor scar from strip harvesting. A Trichophytic closure involves trimming the upper or lower edge of the incision and then closing the wound in such a way that the hair near the edge can grow toward or through the scar. This has the potential to produce a donor scar that is virtually undetectable.⁸⁰

Automation

Short of cloning follicular units in a Petri dish, the future in FUT optimization lies in the development of robotic devices that can harvest the donor strip, isolate individual follicular units, create recipient sites, and insert grafts—all without the risk of human variability or error. A number of creative instruments have been designed toward this end, but thus far their success has been limited.⁸¹⁻⁸⁴ Much of the difficulty lies in the inherent variability of the human scalp and the follicular unit itself. The full automation of a procedure that is labor intensive, organizationally complex, and dependent on the aesthetic judgment of the surgeon remains a formidable challenge.

Hair Cloning

The ability to clone human follicles will revolutionize surgical hair restoration as this will provide an unlimited supply of donor hair. Cloning will potentially allow older patients to enjoy

a full head of hair and younger patients to have their adolescent density and hair line restored. One of the most promising areas of research is Follicular cell implantation (FCI). FCI is based on the ability of the dermal papilla cells, found at the bottom of hair follicles, to interact with the keratinocytes and stimulate new hairs to form. Dermal papilla cells (fibroblasts) can be grown and multiplied in culture, so that a very small number can theoretically produce enough follicles to cover an entire bald scalp.⁸⁵

There a number of obstacles that still need to be overcome before cloning can be a viable option for patients. First, fibroblasts seem to lose their inductive properties when they are cultured over multiple generations; second, the production of cosmetically useful hair – which depends on the interaction between the various cell types – is not yet possible, and third, the research is in its pre-clinical phases, with FDA approval still many years away.

POSTOPERATIVE CARE

Patients are called at home the day after the procedure. Those with a stapled closure are seen in our office 10 days post-op to have alternating staples removed. They return again 7-10 days later to have the remainder of the staples removed. Patients in whom we have used sutures return for only one visit 7 to 10 days post-op.

At 10 to 12 months post-op, all patients return to the office so that we can assess growth, take photos, and discuss the possibility of additional surgery. Those with questions or concerns are seen as frequently as needed. Table 29.12 summarizes typical postoperative instructions.

Table 29.12 Summary of instructions following follicular unit transplantation	
Care of transplanted area	Shampoo every 3-4 hours the day following surgery and then twice daily until the 1-week postoperative visit. Be especially careful when cleaning the transplanted site in the first few days so that the grafts will not be dislodged. However, a thorough but gentle rinsing of the recipient area will minimize crusting and make the transplant less noticeable. When shampooing and rinsing the transplanted area, be gentle for the first 2 weeks following surgery. Do not rub, pick, or scratch, as this may dislodge grafts.
Care of sutured area and non-transplanted parts of scalp	Gently wash the sutured area with your fingertips using a copper peptide-containing shampoo. Shower water may hit the sutured area in the back of your scalp directly. You may use a hair dryer set on warm, but not hot. Do not use any scalp or hair-coloring agents for at least 2 weeks after your procedure.
Postoperative medications	After your procedure, you will be given sleeping medication and medication for pain.
	Itching: You may experience some itching either in the transplanted area or sutured area following your procedure. Hydrocortisone ointment may be applied locally to the areas that itch as needed, up to four times a day.

	Swelling: You will be given an injection of cortisone during your procedure to decrease swelling after surgery. If there is significant swelling, it usually occurs around the second to fifth postoperative day and should not be cause for concern. It resolves by itself after a few days and does not require any special treatment. Cool, wet dressings may be placed over the swollen area, but make sure they do not touch the grafts.
	Bleeding: Before you leave the office, all bleeding will be controlled. Rarely some bleeding may occur after the procedure. If this happens, apply firm continuous pressure on the area. If you are unable to contact us through our pager, go to the nearest emergency room and show our instructions to the physician on duty.
	Exercise, alcohol, smoking and other restrictions: Avoid direct trauma to the head for 2 weeks after the procedure, and abstain from sexual intercourse, alcohol use, and smoking for 3 days after the procedure. After 2 weeks, you may resume your normal daily activities. However, be careful not to stretch the donor area until instructed by your physician.
	Sun: Avoid unprotected exposure to sunlight for 3 months. Wear a hat when you are going to be outside or use a strong sunscreen with an SPF factor (sun protection factor) of 30+.
	Infection: Redness, swelling and slight tenderness are to be expected for the first few days after a procedure. Persistent swelling, pain, or tenderness in the sutured area may be a sign of infection. Fever and/or chills are indications of infection as well. There also may be a discharge or pus in the suture line. If any of these conditions should occur, please contact the office.
	Numbness: Scalp numbness, tingling or similar sensations may occur temporarily. This generally disappears within a few weeks to months, as nerve endings re-grow.
Hair-loss medications	Minoxidil 5%: If you plan to continue this medication, resume it 1 to 2 weeks after the procedure.
	Finasteride 1 mg: This medication should not be discontinued for the surgery.

PITFALLS AND THEIR MANAGEMENT

A number of problems arising in hair transplant surgery have been well described in the literature. These are summarized in Table 29.13, along with their most likely cause, although some etiologies may be multi-factorial. The table also summarizes and methods for preventing and managing them should they occur.^{11,86,87}

Table 29.13 Problems Encountered in Follicular Unit Hair Transplantation and their Management

Problem	Cause	Prevention	Treatment
Syncope	Anxiety, vasovagal reaction	Premedicate with diazepam or atropine, administer local anesthesia with patient lying flat	Lie patient supine and elevate legs
Anxiety, palpitations	β 1- and α -adrenergic effects of epinephrine (adrenaline)	Administer epinephrine slowly, premedicate with diazepam	Patience (the half-life of epinephrine is short); lidocaine for arrhythmias
Hypertension, bradycardia	Epinephrine (adrenaline) interaction with β -blocker	Use selective β 1-blockers	α -adrenergic blockers; treat as cardiac emergency
Agitation, confusion, perioral numbness	Lidocaine toxicity	Use more dilute anesthetic mixture, use ring blocks rather than local infiltration	Intravenous fluids, oxygen, ventilate, Diazepam; treat as cardiac emergency
Pruritus, urticaria, angioedema	Allergic reaction or anaphylaxis to drugs or food	Careful history	Oxygen, epinephrine (adrenaline) subcutaneously
Excessive bleeding	Elevated blood pressure, non-steroidal anti-inflammatory agents	Discontinue antiplatelet agents before surgery, continue antihypertensive medications on day of surgery, pulse oximetry	Operate with patient seated; with elevated blood pressure, consider antihypertensives
Postoperative edema	Surgical trauma, buffered anesthetic	Keep recipient sites small, preoperative and post-operative systemic corticosteroids	Head elevation, additional postoperative corticosteroids

Sterile folliculitis	Growing hair trapped by epidermal overgrowth	Keep grafts slightly elevated	Resolves spontaneously; incision and drainage if persistent
Cyst formation at graft site	Foreign body reaction to imbedded graft	Avoid piggybacking one graft over another in the same site	Incision and drainage, topical and/or systemic antibiotics
Bacterial folliculitis	Picking, scratching, poor hygiene, secondary to infected sterile folliculitis or cyst	Frequent postoperative showering, topical corticosteroids for pruritus	Topical and systemic antibiotics
Infected donor wound	Suture bites too large or tight; with postoperative edema, causes vascular compromise	Harvest a narrower donor strip under less tension, change suturing technique	Systemic antibiotics; if severe, leave portion of donor area to heal by second intention, watch for MSRA ⁹²
Widened scar in donor area	Donor strip too wide or placed too low, patient tendency to heal with stretched scar	Careful history, conservative donor strip located at level of occipital protuberance	Excision ± (can worsen scar), follicular unit extraction
Raised donor scar	History of keloids, racial susceptibility, incision placed too low	Careful history, patient selection, test biopsy	Intralesional corticosteroids
Persistent numbness or paresthesias in the back of the scalp	Transection of branches of the occipital nerves	Limit donor depth to deep subcutaneous layer, use tumescent anesthesia	Subsides spontaneously if the cut nerve branches were small
Hiccups	Possible injury to C2-C4 indirectly stimulating the	Limit donor depth to deep subcutaneous	Chlorpromazine

	vagus or diaphragmatic nerves	layer, use tumescent anesthesia	
Hair texture changes	Trauma to grafts, sebaceous glands removed in dissection	Careful atraumatic dissection and placing, avoid trimming follicular units too close	Usually resolves spontaneously
Skin texture changes	Recipient wounds too large	Make wounds no larger than the equivalent size of a 19-gauge needle at the frontal hairline	Add coverage with one- and two-hair follicular units at the frontal hairline using very small sites
Hair loss in recipient area related to the procedure	Telogen effluvium ⁹³	Possibly effective: small surgical wounds, limit epinephrine, pretreat with finasteride	Terminal non-miniaturized hair should return spontaneously, finasteride
Hair loss in donor area	Strangulation of follicles by sutures, transection of follicles during harvest	Narrower donor strip, meticulous suturing techniques	Follicular unit transplantation, follicular unit extraction, local excision
Failure of transplanted hair to grow	Desiccation, ⁹¹ crush injury, ⁸⁹ transection, ^{90,94,95,96} grafts out of the body too long ⁵²	Keep grafts well hydrated in an isotonic solution, careful microscopic dissection and handling	None

Although FUT eliminates many of the shortcomings of the older techniques, such as a ‘pluggy’ look, a ‘moth-eaten’ donor area, or midline scalp reduction scars, poor aesthetic judgment and techniques that compromise graft growth can still lead to problems. Perhaps because FUT requires large numbers of grafts (using a significant portion of the donor area at one time), because so many staff members are involved in the process, and because some of the problems of small-graft procedures are very difficult to correct, improperly performed follicular unit transplantation can sometimes pose a greater risk to patients than traditional grafting. The problem is compounded by the fact that many physicians perceive FUT as a safe, risk-free procedure and describe it to patients as such.

The remainder of this section will focus on some of the most common mistakes made by FUT practitioners, particularly in the areas of planning, transplant design, and handling large numbers of small grafts. These problems and how they may be avoided are summarized in Table 29.14.

1.	Operating on patients that are too young or prior to trying medical therapy
2.	Failing to identify low donor density prior to surgery
3.	Failing to identify a tight scalp
4.	Harvesting a donor strip that is too wide
5.	Placing the donor incision too low or too high
6.	Using a multi-bladed knife
7.	Crushing grafts during insertion
8.	Allowing grafts to dry
9.	Placing the frontal hairline too far forward
10.	Creating a hairline that is too broad
11.	Angling hair in the wrong direction
12.	Attempting to cover an area that is too large

Operating on patients that are too young or before medical therapy

Patients in their early 20s have their flat adolescent hairline and original density fresh in their memory. A transplant designed with enough frontal and temporal recession to look good for one's entire life will rarely satisfy a younger patient. Creating a density that suits a younger person will not leave enough hair in reserve when their hair loss progresses. In addition, at this age, the extent of future balding is very difficult to anticipate. For these reasons, surgery should rarely be considered in patients with androgenetic alopecia who are younger than 25 years of age.

Often doctors begin medical therapy and schedule surgery at the same time. However, if there is a possibility that using medication, such as finasteride and minoxidil, can make transplantation unnecessary, then the medication should be used for at least a year before any decision on surgery is reached. Medication should be the first line of therapy for all younger patients with androgenetic alopecia, regardless of the degree of their hair loss.

Failing to identify low donor density before surgery

Earlier in this chapter the importance of assessing patient's donor supply with densitometry was stressed. A low donor density, generally less than 1.5 hairs per mm², usually indicates that donor supply is insufficient to create adequate density or coverage and is a relative contraindication to surgery. An exception might be an older person with very conservative goals. High miniaturization in the donor area, particularly in a person under the age of 30, suggests diffuse unpatterned hair loss (DUPA) and is, in this author's opinion, a contraindication to surgery.⁶

Transplanting young patients with low donor density will not only insure a poor cosmetic result, but will also risk a visible scar if the hair is worn short. FUE, which is sometimes considered in such cases, is not appropriate because it further limits the total available hair. In fact, because the contrast between bald and non-balding scalp in patients with low donor density is naturally low, their best option tends to be wearing their hair short, to decrease the contrast further (rather than having surgery).

Failing to identify a tight scalp

Assessing scalp laxity is an underappreciated aspect of the patient evaluation, probably because it is difficult to quantify. However, a tight scalp severely limits the total amount of harvestable donor hair and can constitute a contraindication to surgery, except when patients have extremely conservative goals or are expected to experience only limited balding. The constraints that poor scalp laxity impose generally manifest themselves after the first transplant session. Though laxity should be judged in the preoperative evaluation, the intraoperative assessment, made while suturing, is most accurate in predicting future difficulties. Therefore, every operative report should include a record of the ease of closure and intraoperative suture tension.⁶

A tight scalp can be loosened by placing the palm of the hands on the back/sides of the head with the fingers pointing upwards and thumbs pointing towards each other. The scalp is then moved up and down as vigorous as possible using the palms and thumbs, with the fingers resting stationary on the top of the scalp. The exercise should be performed for a total of 15 minutes every evening at bedtime. One should try to begin the exercise at least 8 weeks prior to surgery and continue the exercise up to the day before the procedure.⁸⁸

Harvesting a donor strip that is too wide

In large sessions, it can be tempting to take a slightly wider donor strip in order to conserve on length. A strip that is 25 cm by 1 cm, for instance, can be shortened by 6 cm if widened by just 3 mm and yield the same amount of hair. However, a wide strip puts unnecessary tension on the donor closure and is probably the most common iatrogenic cause of widened scars. If larger sessions are appropriate, and the scalp lacks adequate mobility, the surgeon should consider a longer incision rather than a wider one.

If a wide donor strip has been identified as the likely cause of a stretched scar, it is advisable to wait at least 8 months, to give the scar a chance to mature and regain some of its original laxity. When the next excision is made the strip should measure at least 3–6 mm narrower than the previous one. Attempts to remove the entire width or length of the old scar invariably lead to a recurrence, or worsening, of the old scar. To facilitate healing, the new excision should extend to the patient's hair-bearing edge, and be significantly shorter than the old incision.

Unfortunately, attempts to re-excise scars commonly result in either no improvement or an even wider defect. For this reason, we have been using follicular unit extraction to place hair directly into the scar as a primary treatment. It has been our experience, however, that transplanted hair will not grow in a thickened scar, therefore, any hypertrophic scars should be thinned with intralesional corticosteroids prior to implantation and then a small test area should be transplanted

first, to see if the hair will grow and to make sure that the scar doesn't thicken from the grafting.⁷⁴

Placing the donor incision too low

The ideal position for the donor incision is in the mid-portion of the permanent zone that lies, in most people, at the level of the external occipital protuberance and the superior nuchal line. The muscles of the neck insert into the inferior portion of this ridge, so an incision below this anatomic landmark will be impacted by the muscle movement beneath it. A stretched scar in this location is extremely difficult to repair because re-excision, even with undermining and layered closure, tends to heal with a wider scar. In addition, one is more likely to cut through fascia with a low donor incision; and once the fascia has been violated, the risk of widening the scar rises considerably.⁴⁰ To compound the problem, some patient's nuchal hair thins dramatically over time rendering low donor scars visible.

Harvesting hair too high in the donor area risks lack of permanence of the transplanted hair because this hair may be subject to androgenic alopecia. The donor scar may also become visible if the donor fringe narrows further. In addition, a high donor scar has a greater risk of a widening compared to one placed at the level of the occipital protuberance. Incisions made too high are best left untreated, unless poor surgical technique has been identified as the cause. The temptation to transplant permanent donor hair into a high scar should be done with caution, as progressive balding may isolate the hair-bearing scar, presenting new cosmetic problems.

In the case of young patients with traumatic scars and hair-loss patterns that are still unclear, follicular unit extraction can function as a hedge against this risk. If the hair is harvested from the immediate vicinity of the scar, any future balding will affect the transplanted hair in the scar at the same rate as the hair surrounding it.

Using a multi-bladed knife

In order to save time, doctors performing large transplants may use a multi-bladed knife (one with three or more blades) for harvesting donor tissue. The resulting pre-sliced multiple, thin strips are much easier to work with than a single intact strip. Unfortunately, harvesting this way causes unacceptable levels of follicular transection and destroys naturally occurring follicular units. This technique is incompatible with Follicular Unit Transplantation.²⁷

Crushing grafts during insertion

Proper placing technique necessitates the use of forceps to grasp the graft by the fat below the bulb or by the dermis alongside the hair shaft in order to avoid damaging the germinative components of the follicle. Though placers often exercise enormous care while initially grasping the graft, there is a tendency to become rougher when repositioning the forceps for further inserting, replacing a popped graft or transferring grafts from the holding solution to the fingers. As follicular units and other small grafts are particularly susceptible to crush injury, improper handling can more than negate the benefits of careful stereomicroscopic dissection.^{89,90}

Allowing grafts to dry

A study using electron microscopy, has shown that desiccation is by far the most significant form of injury to grafts and that this makes them much more susceptible to other forms of injury, such as a mechanical trauma and warming. Grafts should therefore be kept well-hydrated with chilled isotonic solution (such as Ringer's lactate) from the moment the tissue is harvested until the time they are reinserted into the scalp.⁹¹

Placing the frontal hairline too far forward (too low)

Despite the fact that individual follicular units at the hairline in themselves look natural, their proper placement is no less important than in traditional grafting. The frontal hairline should be placed no lower than 1.5 cm above the upper brow crease.^{1,7} If the underlying skin is normal, follicular units placed too low can be removed with an alexandrite (755 nm) or diode (800 nm, 810 nm) laser. Electrolysis is more difficult and time-consuming with transplanted follicles, but should also be considered. Punch excision is too imprecise for very small grafts and FUE, with wounds left to heal by secondary intention, risks visible scarring.

Creating a hairline that is too broad

Since significant temporal recession is characteristic of a normal adult man's hairline, a broad, flat transplanted hairline will not age well and can cause cosmetic problems if baldness becomes extensive. The treatment is the same as with low hairlines, but if grafts larger than follicular units were used, and/or if there is scarring of the recipient skin, punch excision with reutilization of the hair should be considered.

Angling hair in the wrong direction

As noted earlier, in the front and top part of the scalp, hair grows in a distinctly forward direction, changing to a radial pattern as it approaches the crown. Hair emerges from the scalp at an acute angle, with the hair lying practically flush to the skin at the temples.

There has been a tendency among some hair restoration surgeons to transplant grafts perpendicular to the skin—probably due to the fact that the mechanics of the old plug procedures made sharp angling technically difficult. The cosmetic consequence of this is most apparent at the frontal hairline. When the hair is perpendicular, the viewer's eye is guided to the base of the hair shaft where it inserts into the skin; conversely, when hair is transplanted in its natural, forward-pointing position, it is bowed by grooming and the eye settles on the body of the hair shaft.

When grafts at the frontal hairline are transplanted in a radial direction, combing the hair in any style becomes problematic as the transplanted hair fans outward and becomes unmanageable. When the hair is pointed forward it can be groomed as a single unit increasing the illusion of fullness. As with low or broad hairlines, hair that is angled in the wrong direction, particularly in the frontal hairline, should often be removed rather than camouflaged.

Attempting to cover an area that is too large

Attempting to cover an area greater than the donor supply can adequately fill may leave cosmetically important areas thin or untransplanted. As a general guide the first region of the scalp to bald is generally the area where you should be most hesitant to transplant. Recession at the temples and thinning in the crown are usually the earliest manifestations of baldness, but they are acceptable, especially as a person ages, so these areas may be left untransplanted. The central forelock region, however, is generally late to bald, but when balding occurs the patient loses the frame to his face and its restoration becomes essential.⁶

Whether or not these areas need coverage at the time of the initial transplant, an adequate amount of hair must always be reserved for critical areas, such as the forelock and top of the scalp. If donor reserves are limited, the transplantation of less critical areas should be postponed or avoided all together.⁶

SUMMARY

Developed within the past decade, follicular unit transplantation has emerged as the ‘gold standard’ in surgical hair restoration. In conserving donor hair, achieving optimal coverage, and creating a natural look, FUT represents a significant advance over earlier methods of hair restoration. Appropriately, it also demands considerably more from its practitioners. Surgical teams must develop the skill and stamina for the delicate handling of large numbers of follicular unit grafts, while surgeons must cultivate a keen aesthetic sensibility with regard to transplant design and graft placement.

In view of the psychological aspects of hair loss, follicular unit transplantation requires a thorough preoperative assessment to understand the patient’s expectations, a careful examination to determine if surgery is appropriate and, most importantly, the establishment of realistic goals. If the surgical route is chosen, meticulous attention to detail is required in every aspect of the procedure, so that these goals may be realized. It is a daunting task for the hair restoration surgeon and team to develop the necessary expertise for perfecting follicular unit transplantation; but when they do the procedure can benefit patients for their lifetime.

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